

Ensembles of Radial Basis Function Networks for Spectroscopic Detection of Cervical Precancer

Kagan Tumer, *Member, IEEE*, Nirmala Ramanujam, Joydeep Ghosh, and Rebecca Richards-Kortum*

Abstract—The mortality related to cervical cancer can be substantially reduced through early detection and treatment. However, current detection techniques, such as Pap smear and colposcopy, fail to achieve a concurrently high sensitivity and specificity. *In vivo* fluorescence spectroscopy is a technique which quickly, noninvasively and quantitatively probes the biochemical and morphological changes that occur in precancerous tissue. A multivariate statistical algorithm was used to extract clinically useful information from tissue spectra acquired from 361 cervical sites from 95 patients at 337-, 380-, and 460-nm excitation wavelengths. The multivariate statistical analysis was also employed to reduce the number of fluorescence excitation-emission wavelength pairs required to discriminate healthy tissue samples from precancerous tissue samples. The use of connectionist methods such as multilayered perceptrons, radial basis function (RBF) networks, and ensembles of such networks was investigated. RBF ensemble algorithms based on fluorescence spectra potentially provide automated and near real-time implementation of precancer detection in the hands of nonexperts. The results are more reliable, direct, and accurate than those achieved by either human experts or multivariate statistical algorithms.

Index Terms—Cancer, fluorescence spectroscopy, multivariate statistical analysis, neural network.

I. INTRODUCTION

CERVICAL carcinoma is the second most common cancer in women worldwide, exceeded only by breast cancer [1]. The mortality related to cervical cancer can be reduced if this disease is detected at its precancerous state, known as squamous intraepithelial lesion (SIL) [2]. Even though widespread use of organized screening (Pap smear) and diagnostic (colposcopy) programs are currently in place, approximately 15 900 new cases of cervical cancer and 4900 cervical cancer-related deaths were reported in the United States alone, in 1995 [1].

Currently, the primary screening tool for the detection of cervical cancer and its precursor is the Pap smear [3]. In a

Manuscript received April 26, 1997; revised February 23, 1998. This research was supported in part by the National Science Foundation (NSF) under Grant ECS 9307632, by the Air Force Office of Scientific Research (AFOSR) under Contract F49620-93-1-0307, and by Lifesplex, Inc., Kirkland, WA. Asterisk indicates corresponding author.

K. Tumer is with Caelum Research, NASA Ames Research Center, Moffett Field, CA 94035-1000 USA.

N. Ramanujam is with the Department of Biochemistry and Biophysics, Johnson Research Foundation, University of Pennsylvania, School of Medicine, Philadelphia, PA 19104 USA.

J. Ghosh is with the Department of Electrical and Computer Engineering, The University of Texas at Austin, Austin, TX 78712 USA.

*R. Richards-Kortum is with the Biomedical Engineering Program, The University of Texas at Austin, Austin, TX 78712-1084 USA (e-mail: kortum@mail.utexas.edu).

Publisher Item Identifier S 0018-9294(98)05327-0.

Pap test, a large number of cells obtained by scraping the cervical epithelium are smeared onto a slide which is then fixed and stained for cytologic examination. Each smear is then examined under a microscope for the presence of neoplastic cells [4]. The Pap smear is unable to achieve a concurrently high sensitivity¹ and high specificity² [5]. The accuracy of the Pap smear is limited by both sampling and reading errors [6]. Approximately 60% of false-negative smears are attributed to insufficient sampling; the remaining 40% are due to reading errors. Because of the monotony and fatigue associated with reading Pap smears (50 000–300 000 cells per slide), the American Society of Cytology has proposed that a cyto-technologist should be limited to evaluating no more than 12 000 smears annually [7]. As a result, accurate Pap-smear screening is labor intensive and requires highly trained professionals. Some new tools (Thinkprep, Papnet, and Autopap) to assist cyto-technologists have recently been introduced, but they are all based on invasive techniques [8].

A patient with a Pap smear interpreted as indicating the presence of SIL is followed up by a diagnostic procedure, called colposcopy [3]. During a colposcopic examination, the cervix is stained with acetic acid and viewed through a low-power microscope to identify potential precancerous sites. Subsequently, suspicious sites are biopsied and then histologically examined to confirm the presence, extent, and severity of the lesion [9]. Colposcopic examination in expert hands maintains a high sensitivity at the expense of a significantly low specificity, leading to many unnecessary biopsies [10]. In spite of the poor specificity of this technique, extensive training is required to achieve this skill level. Furthermore, since this procedure involves biopsy, which requires histologic evaluation, diagnosis is not immediate [3].

Laser-induced fluorescence spectroscopy is an optical technique which has the capability to quickly, noninvasively, and quantitatively probe the biochemical and morphological changes that occur as tissue becomes neoplastic. The altered biochemical and morphological state of the neoplastic tissue is reflected in the spectral characteristics of the measured fluorescence. This spectral information can be correlated to tissue histopathology, the current “gold standard” to develop clinically effective screening algorithms. These mathematical algorithms can be implemented in software, potentially enabling automated, fast, noninvasive, and accurate precancer

¹Sensitivity is the correct classification percentage on the precancerous tissue samples.

²Specificity is the correct classification percentage on normal tissue samples.

detection in hands of nonexperts. Although a complete understanding of the quantitative information contained within a tissue fluorescence spectrum has not been achieved, many groups have investigated the use of fluorescence spectroscopy for real-time, noninvasive, automated characterization of tissue pathology [11]–[17].

A detection technique for human cervical precancer based on laser-induced fluorescence spectroscopy has been developed recently [18]. Discrimination was achieved using a multivariate statistical algorithm (MSA) based on principal component analysis (PCA) and logistic discrimination of tissue spectra acquired *in vivo*. This linear method of algorithm development demonstrated an improved classification accuracy relative to both the Pap smear and colposcopy in expert hands. In this paper, we investigate neural network-based nonlinear methods for algorithm development, and compare them to both the MSA and conventional clinical methods. Specifically, we investigate the performance of multilayered perceptron (MLP) and radial basis function (RBF) networks, and ensembles of these networks, on cervical tissue fluorescence spectra. The connectionist methods aim at improving the classification accuracy and reliability of the MSA, as well as simplifying the decision making process. Section II presents the data collection/processing techniques. In Section III, the MSA, and the neural network-based methods are described. Section IV contains the results of our analysis, and compares the neural network results to that of the MSA and current clinical detection methods. A discussion of the results is given in Section V.

II. DATA COLLECTION AND PROCESSING

A. Instrumentation and Clinical Measurements

A portable fluorimeter consisting of two nitrogen pumped-dye lasers, a fiber-optic probe and a polychromator coupled to an intensified diode array controlled by an optical multichannel analyzer was utilized to measure fluorescence spectra from the cervix *in vivo* at three excitation wavelengths: 337, 380, and 460 nm [18]. Data acquisition, calibration, and processing have been described in detail elsewhere [18].

A randomly selected group of nonpregnant patients referred to the colposcopy clinic of the University of Texas MD Anderson Cancer Center on the basis of abnormal cervical cytology was asked to participate in the *in vivo* fluorescence spectroscopy study. Informed consent was obtained from each patient who participated and the study was reviewed and approved by the Institutional Review Boards of the University of Texas, Austin, and the University of Texas, MD Anderson Cancer Center.

Each patient underwent a complete history and a physical examination including a pelvic exam, a Pap smear, and colposcopy of the cervix, vagina, and vulva. After colposcopic examination of the cervix, but before tissue biopsy, fluorescence spectra were acquired on average from two colposcopically abnormal sites, two colposcopically normal squamous sites, and one normal columnar site (if colposcopically visible) from each patient, from a total of 361 cervical sites in 95 patients.

TABLE I
HISTOPATHOLOGIC CLASSIFICATIONS OF
SAMPLES FROM THE TRAINING AND TEST SETS

Histo-pathology	Target Classification	Training Set	Test Set
Normal Squamous	non-SIL	94	94
Normal Columnar		13	14
Inflammation		15	14
LG SIL	SIL	23	24
HG SIL		35	35

Note, biopsies for histological evaluation were not obtained from colposcopically normal squamous and columnar tissue sites to comply with routine patient care procedure.

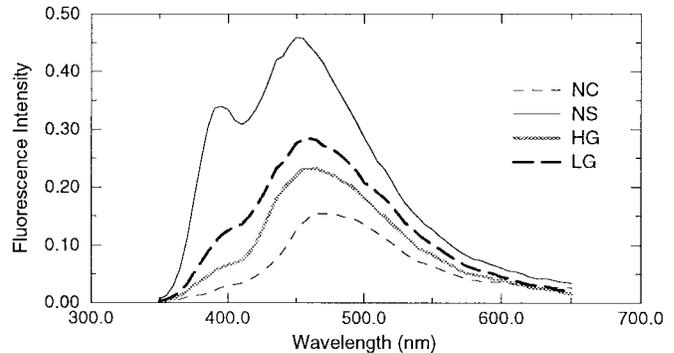


Fig. 1. Fluorescence spectra from a typical patient at 337-nm excitation.

Tissue biopsies were obtained only from abnormal sites identified by colposcopy and, subsequently, analyzed by the probe to comply with routine patient care procedure. All tissue biopsies were fixed in formalin and submitted for histologic examination. Hemotoxylin and eosin-stained sections of each biopsy specimen were evaluated by a panel of four board certified pathologists and a consensus diagnosis was established using the Bethesda classification system [2]. This classification system which has previously been used to grade cytologic specimens has now been extended to classification of histology samples. Samples were classified as normal squamous (NS), normal columnar (NC), inflammation,³ low-grade (LG) SIL and high-grade (HG) SIL. Table I provides the number of samples in the training (calibration) and test (prediction) sets.

B. Spectral Data

Fig. 1 illustrates average fluorescence spectra per site acquired from cervical sites at 337-nm excitation from a typical patient. All fluorescence intensities are reported in the same set of calibrated units. Evaluation of the spectra at 337-nm excitation indicates that the fluorescence intensity of SIL's (LG and HG) is less than that of the corresponding normal squamous tissue; however, their fluorescence intensity is greater than that of the corresponding normal columnar tissue over the entire emission spectrum.

³In this paper, we will not focus on the classification of tissues with inflammation. Evaluation of these tissues using both MSA and neural networks indicates that they are nearly indistinguishable from SIL's based on the spectral data presented here [19], [20]. To remedy the situation, different optical spectroscopic techniques are needed.

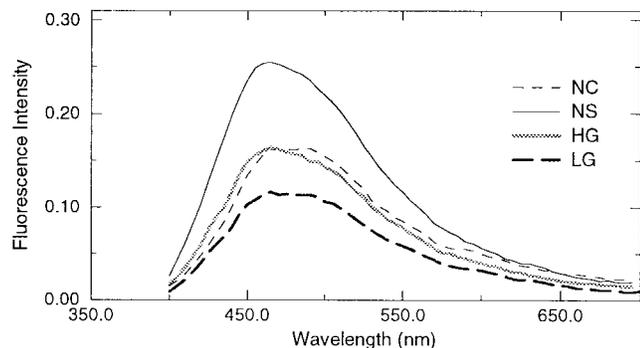


Fig. 2. Fluorescence spectra from a typical patient at 380-nm excitation.

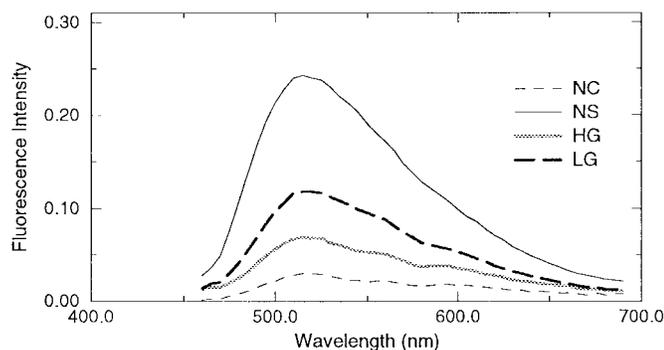


Fig. 3. Fluorescence spectra from a typical patient at 460-nm excitation.

Fig. 2 illustrates average fluorescence spectra per site acquired from cervical sites at 380-nm excitation, from the same patient. In Fig. 2, the fluorescence intensity of SIL's is less than that of the corresponding normal squamous tissue, with the LG SIL exhibiting the weakest fluorescence intensity over the entire emission spectrum. Note that the fluorescence intensity of the normal columnar sample is indistinguishable from that of the HG SIL. Fig. 3 illustrates spectra at 460-nm excitation from the same patient. Evaluation of Fig. 3 indicates that the fluorescence intensity of SIL's is less than that of the corresponding normal squamous tissue and greater than that of the corresponding normal columnar sample over the entire emission spectrum.

Tissue fluorescence spectra at 337-nm excitation consist of intensities at a total of 59 emission wavelengths; tissue spectra at 380-nm excitation consist of intensities at a total of 56 emission wavelengths and that at 460-nm excitation consist of intensities at 45 emission wavelengths. Hence, fluorescence spectra at all three excitation wavelengths comprise of fluorescence intensities at a total of 160 excitation-emission wavelength pairs.

III. ALGORITHM DEVELOPMENT

In this section, the development of the multivariate statistical algorithm and the neural network-based algorithms are described. Each type of algorithm was utilized to develop a detection method that can effectively discriminate between SIL's and non-SIL's (normal squamous and normal columnar).

A. Multivariate Statistical Algorithms

1) *Full-Parameter Multivariate Statistical Algorithm*: The MSA development described in [18] consists of the following five steps:

- 1) preprocessing to reduce inter-patient and intra-patient variation of fluorescence spectra from a tissue type, using either normalization or normalization followed by mean-scaling;
- 2) dimension reduction of the preprocessed tissue fluorescence spectra using PCA;
- 3) selection of diagnostically relevant PC's, using an unpaired, one-sided student's *t*-test;
- 4) development of a classification algorithm based on logistic discrimination, using the diagnostically relevant PC's;
- 5) retrospective and prospective evaluation of the algorithm's accuracy on a training and test set, respectively.

This process of algorithm development was applied to tissue fluorescence spectra acquired at all three excitation wavelengths, as described in detail in [18]. Discrimination between the SIL's and the two normal tissue types was achieved using a composite algorithm of two independently developed *constituent* algorithms. Constituent algorithm (1), which is based on tissue spectra that have been preprocessed by normalization, discriminates between SIL's and normal squamous tissue samples. Constituent algorithm (2), which is based on tissue spectra that have been preprocessed by normalization followed by mean-scaling, discriminates between SIL's and normal columnar tissue samples. The classification outputs from both constituent algorithms were used to determine whether a sample being evaluated is SIL/non-SIL. A sample is first presented to constituent algorithm (1). If it is classified as non-SIL, the algorithm terminates. If it is classified as SIL, then the sample is presented to constituent algorithm (2), and the result of that algorithm determines the final classification of the tissue sample. Fig. 4 illustrates this procedure.

2) *Reduced-Parameter Multivariate Statistical Algorithms*: The full-parameter MSA utilizes fluorescence spectra at all three excitation wavelengths to develop a classification scheme for cervical precancer detection; the fluorescence spectra at these three excitation wavelengths correspond to fluorescence intensities at a total of 160 excitation-emission wavelength pairs (5-nm spectral resolution). However, there is a significant cost penalty for using all 160 values. To alleviate this concern, a more cost-effective fluorescence imaging system can be developed, if the number of required excitation-emission wavelength pairs at which fluorescence intensities need to be recorded is significantly reduced. For example, if the number of required excitation-emission wavelength pairs that need to be measured can be reduced by an order of magnitude, the polychromator and intensified diode array can be replaced by a mechanical filter assembly and a single-channel detector. The resulting system represents a substantial decrease in cost and complexity of this instrumentation.

We have shown that component loadings calculated from PCA [18] can be used to reduce the number of fluorescence

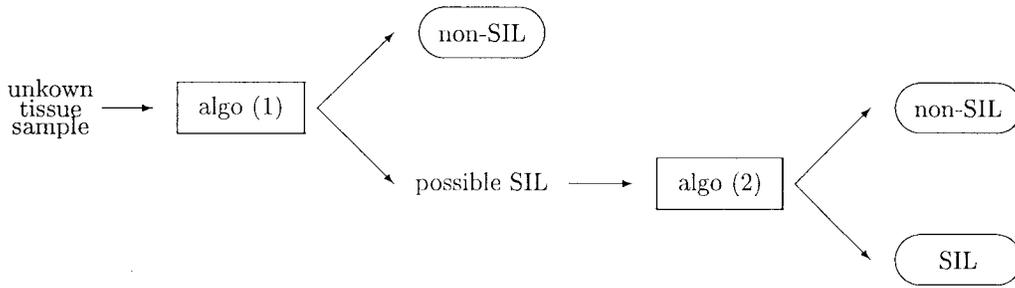


Fig. 4. Schematic representation of the composite MSA algorithm.

TABLE II
FLUORESCENCE INTENSITIES AT 13 EXCITATION-EMISSION WAVELENGTH
PAIRS NEEDED TO REDEVELOP THE TWO REDUCED-PARAMETER
CONSTITUENT ALGORITHMS. ASIDE FROM THE PREPROCESSING, THE
TWO SETS ONLY DIFFERED IN TWO SELECTIONS, IDENTIFIED BY *

Features for Algorithm (1) (Normalized)	Features for Algorithm (2) (Normalized and Mean Scaled)
$\lambda_{ex}, \lambda_{em}$	$\lambda_{ex}, \lambda_{em}$
337, 410 nm	337, 410 nm
337, 430 nm	337, 430 nm
337, 510 nm	337, 510 nm
337, 580 nm	337, 580 nm
380, 410 nm	380, 410 nm
380, 430 nm	380, 430 nm
380, 510 nm	380, 510 nm
380, 580 nm	380, 580 nm
380, 640 nm	380, 600 nm *
460, 580 nm	460, 580 nm
460, 600 nm	460, 600 nm
460, 620 nm	460, 620 nm
460, 640 nm	460, 660 nm *

excitation-emission wavelength pairs required to generate the constituent algorithms from 160 to 13, with a minimal decrease in classification accuracy. The component loadings represent the correlation between a PC and the preprocessed fluorescence spectra at each excitation-emission wavelength pair. More precisely, the diagnostically relevant PC's of the full-parameter set (160 variables) selected using the Student's t -test are used to calculate the component loadings. The intensity/wavelength pairs that show a strong correlation to the relevant PC's form the reduced-parameter set. Table II shows the fluorescence intensities at the reduced number of excitation-emission wavelength pairs. These pairs are used to redevelop constituent algorithms (1) and (2) using the MSA process.

B. Algorithms Based on Neural Networks

The second stage of algorithm development consists of evaluating the applicability of neural networks to this problem. For this study, we consider two types of feed-forward neural networks, the MLP and the RBF network [21].

1) *Multilayered Perceptrons*: The MLP, probably the most commonly used neural network architecture, is a feed-forward neural network with an input layer, an output layer, and, possibly, multiple hidden layers. Each layer is connected only to the subsequent layer by variable weights which are adjusted

to minimize a predetermined cost function, such as the mean square error (MSE). For an MLP with one hidden layer, the responses of the k th hidden unit, h_k , and the j th output, o_j , are, respectively, given by

$$h_k = g\left(\sum_i v_{ki}x_i\right) \quad (1)$$

and

$$o_j = f\left(\sum_k w_{jk}h_k\right) \quad (2)$$

where the input to hidden connections strength are given by v , the hidden to output connection strengths are given by w , and indices i and k sum over the input (x) and hidden units respectively. The activation functions $f(\cdot)$ and $g(\cdot)$ are sigmoidal functions.

In order to adapt the weights, the *backpropagation* algorithm is generally used. The principle of this algorithm is based on distributing the "blame" or the contribution of each unit to the overall error, across the proper weights. Further details can be found in [22] and [23]. In this study, we only explore MLP's with a single hidden layer.

2) *Radial Basis Function Networks*: RBF networks are feed-forward networks with a single hidden layer where the activation function is a radially symmetric basis function. The output units perform a weighted sum over the outputs of the hidden units (also called kernels). One class of RBF's that is of particular interest consists of those with Gaussian kernels, or where the basis' ($R_j(\mathbf{x})$) have the following activation function [10]:

$$R_j(\mathbf{x}) = e^{-(1/2)\left(\|\mathbf{x}-\mathbf{x}_j\|^2/\sigma_j^2\right)} \quad (3)$$

where σ_j determines the width of the receptive field and \mathbf{x}_j determines the centroid of the j th kernel, respectively. The j th hidden node has a maximum output of one when input $\mathbf{x} = \mathbf{x}_j$. Important parameters in the design of RBF networks include the number, location, and receptive field widths of the kernels.

3) *Combining Multiple Networks*: The performance of a given MLP or RBF network depends on many parameters, including size, learning rate, training strategy, and initial weights. These differences result in different classification decisions, making the selection of a single "best" network a delicate matter. This problem is further compounded when the amount of training patterns is limited, because the definition of "best" depends on the particular validation set chosen. In

such cases, it is difficult to ascertain whether one network will outperform all others given a different test set, as the validation sets are small. Furthermore, selecting only one classifier discards a large amount of potentially relevant information. In order to use all the available data, and to increase both the performance and the reliability of the methods, one can pool the outputs of the individual classifiers before a classification decision is made [24]–[29].

In this study, we use the median combiner, f_i^{med} , which belongs to the class of order statistics combiners introduced in [30], and the averaging combiner, f_i^{ave} , which performs an arithmetic average of the corresponding outputs

$$f_i^{\text{med}}(x) = \begin{cases} \frac{f_i^{(N/2):N}(x) + f_i^{(N/2)+1:N}(x)}{2}, & \text{if } N \text{ is even} \\ f_i^{(\frac{N+1}{2}):N}(x), & \text{if } N \text{ is odd} \end{cases} \quad (4)$$

and

$$f_i^{\text{ave}}(x) = \frac{1}{N} \sum_{m=1}^N f_i^m(x) \quad (5)$$

where N represents the number of available classifiers, and $f_i^m(x)$ the i th output of the m th classifier for input x . We selected these two combiners, because of their simplicity in both interpretation and implementation, and because they typically result in good and robust performance [30], [31].

IV. RESULTS

In this section we discuss the application of various neural network-based algorithms to the spectral data discussed in Section II. In order to establish the validity of using the neural network ensembles to perform this task, we conducted the following sets of experiments:

- 1) To evaluate the suitability of the neural network methods, we redeveloped constituent algorithms (1) and (2) with the RBF and MLP ensembles using the following:
 - a) the full-parameter, preprocessed data sets;
 - b) the diagnostically relevant PC's obtained from the full-parameter, preprocessed data sets;
 - c) the reduced-parameter, preprocessed data sets.
- 2) Reduce the two-step algorithm developed using MSA into a single-step algorithm using RBF and/or MLP ensembles.
- 3) Produce full comparative results showing the neural network ensembles' classification accuracy and reliability relative to the MSA, Pap smear and colposcopy.

A. Neural Network Ensembles on Full-Parameter Set

The first step in applying the neural network ensembles to this problem consisted of determining whether the algorithms were suited for this task. To that end, both the MLP and RBF ensembles were used to separate the normal squamous tissues from the SIL's [constituent algorithm (1)]. The task proved to be impossible using the full 160 parameters, mainly because of the small number of training samples. Constraining the number of weights required to handle 160 parameters with the available data resulted in a highly ill-posed problem [32].

TABLE III
ACCURACY OF *constituent* ALGORITHM (1) FOR DIFFERENTIATING BETWEEN SIL'S AND NORMAL SQUAMOUS TISSUES, BASED ON THE DIAGNOSTICALLY RELEVANT PC'S OF THE FULL-PARAMETER SET CONTAINING NORMALIZED SPECTRA. ALL RESULTS ARE BASED ON TEST TEST PERFORMANCE

Algorithm	Specificity	Sensitivity
MSA	68%	88%
RBF-single	71% \pm 3.8%	86% \pm 2.1%
MLP-single	65% \pm 0.0%	83% \pm 1.1%
RBF-ave	74% \pm 0.7%	86% \pm 0.5%
RBF-med	72% \pm 1.7%	86% \pm 1.5%
MLP-ave	65% \pm 0.0%	83% \pm 0.0%
MLP-med	65% \pm 0.0%	84% \pm 0.7%

As a result, the network tried to memorize the training data and performed poorly on the test set. In order to avoid this pitfall, we used the three diagnostically relevant PC's from the full-parameter data set containing normalized fluorescence spectra. (Note, the MSA was developed using these same three PC's, because it also was unable to solve this problem using the full-parameter data.) Both networks had two outputs, each representing the posterior probabilities of the corresponding class. The MLP networks had a single hidden layer with three hidden units, and the RBF networks had three kernels which were initialized by a k -means algorithm on the training set.⁴

The ensemble results reported in Table III correspond to the pooling of 20 different classifiers (MLP's or RBF's), each of which started from different random weight initializations, using the average and median combiners, respectively.⁵ All the results reported in this paper are based on test set performance. We report the sensitivity and specificity values separately, rather than a single classification rate, to emphasize the difference between a false-positive and a false-negative. For an application such as precancer detection, the cost of a misclassification varies greatly from one class to another. Erroneously labeling a healthy tissue as precancerous can be corrected when further tests are performed. Labeling a precancerous tissue as healthy, however, can lead to disastrous consequences. Therefore, for algorithm (1), we increased the cost of a misclassified SIL until the sensitivity⁶ reached a satisfactory level (comparable to the sensitivity of current clinical detection).

For this experiment, the RBF-based combiners provide higher specificity than either the MLP ensembles or the MSA, for a similar sensitivity. This experiment was conducted to ensure that neural network ensembles could duplicate the MSA results [18], using the PC's extracted from the full-parameter set.

B. Neural Networks on Reduced-Parameter Set (Two-Step Algorithm)

The second step in applying neural networks to this problem consisted of determining whether the neural network ensemble

⁴The appropriate sizes of both the MLP and the RBF network were determined experimentally. Because we found that the performance was comparable over fairly large range of network sizes, it was not necessary to use sophisticated methods.

⁵The gains due to combining were minimal if more classifiers were used.

⁶In the results reported in this section, the cost of misclassifying a SIL was two times the cost of misclassifying a normal tissue sample.

performance on the reduced-parameter data set was acceptable, by retracing the development of the two-step process outlined for the MSA. Specifically, the neural network ensemble was applied to the preprocessed reduced-parameter data sets, used to develop constituent algorithms (1) and (2).

For *constituent* algorithm (1), the RBF kernels were initialized using a k -means clustering algorithm on the training set containing normal squamous tissue samples and SIL's. The RBF networks had ten kernels, whose locations and spreads were adjusted during training. The MLP had one hidden layer with ten hidden units. For *constituent* algorithm (2), we selected ten kernels, half of which were fixed to patterns from the columnar normal class, while the other half were initialized using a k -means algorithm. Neither the kernel locations nor their spreads were adjusted during training. This process was adopted to rectify the large discrepancy between the number of samples from each category (13 for columnar normal versus 58 for SIL's). In this case, the MLP algorithm had ten hidden units. Training was stopped when the performance on the training set slowed down sufficiently to suggest further training would cause overtraining.⁷

Once a stopping time was selected, 20 cases were run for each algorithm.⁸

The ensemble results reported are based on the pooling of 20 different runs of RBF networks, initialized and trained as described in the previous section. Once again we increased the cost of misclassifying a SIL in order to increase the sensitivity at the expense of reducing the specificity. For algorithm (1), the cost of a misclassified SIL was 2.5 times the cost of a misclassified normal tissue sample. The sensitivity and specificity values for *constituent* algorithm (1) based on MSA, MLP, and RBF ensembles are provided in Table IV. Table V presents sensitivity and specificity values for *constituent* algorithm (2) obtained from MSA, and MLP and RBF ensembles. In this case, there was no need to increase the cost of a misclassified SIL for the RBF network, because of the high prominence of SIL's in the training set. For the MLP's however, the cost of normal columnars had to be increased in order to obtain classification decisions.⁹

For both algorithms (1) and (2), the RBF-based combiners provide higher specificity than the MSA, while the MLP ensemble does so only for algorithm (1). Furthermore, in the case of the RBF algorithm, this increase is achieved without a decrease in the sensitivity. The median combiner provides results similar to those of the average combiner, except for algorithm (2) where it provides better specificity. From these experiments we can conclude that the RBF network is better suited for this task than the MLP. While in some other work, such as classification of sonar signals, we have found combining MLP's with RBF's to be fruitful [33], in this problem we were surprised to find that combining MLP's and

⁷In general, leave-one-out or K-fold cross validation would be more desirable. However, because the results were similar over a large window of stopping times, this more naive method seemed adequate.

⁸Each run has a different initial set of kernels/spreads/weights.

⁹When the cost of a misclassified SIL was the same as the cost of a misclassified normal columnar, all patterns were classified as SIL's by the MLP. Only when the cost of normal columnars was increased did the MLP start to make nontrivial classification decisions.

TABLE IV
ACCURACY OF *constituent* ALGORITHM (1) FOR DIFFERENTIATING SIL'S AND NORMAL SQUAMOUS TISSUES, BASED ON THE REDUCED-PARAMETER SET CONTAINING NORMALIZED SPECTRA

Algorithm	Specificity	Sensitivity
MSA	63%	90%
RBF-single	66% \pm 2.7%	84% \pm 2.0%
MLP-single	61% \pm 0.0%	91% \pm 0.2%
RBF-ave	66% \pm 0.6%	90% \pm 0.0%
RBF-med	66% \pm 1.1%	90% \pm 0.7%
MLP-ave	61% \pm 0.0%	91% \pm 0.0%
MLP-med	61% \pm 0.0%	91% \pm 0.0%

TABLE V
ACCURACY OF *constituent* ALGORITHM (2) FOR DIFFERENTIATING SIL'S AND NORMAL COLUMNAR TISSUES, BASED ON THE REDUCED-PARAMETER SET CONTAINING NORMALIZED AND MEAN-SCALED SPECTRA

Algorithm	Specificity	Sensitivity
MSA	36%	97%
RBF-single	35% \pm 15%	97% \pm 1.6%
MLP-single	47% \pm 6.9%	89% \pm 3.5%
RBF-ave	37% \pm 5.0%	97% \pm 0.0%
RBF-med	44% \pm 7.5%	97% \pm 0.0%
MLP-ave	50% \pm 0.0%	88% \pm 0.7%
MLP-med	50% \pm 0.0%	89% \pm 2.5%

RBF's always gave worse results than combining RBF's alone. We hypothesize that the fine tuning required to accommodate the varying number of class samples poses severe problems for MLP's global computations. The RBF networks significantly alleviate this problem by placing the kernels in the appropriate places.

We conducted this experiment to not only demonstrate that the reduced-parameter data set does not compromise the performance of the network ensembles, but also to compare the RBF and MLP ensembles. Having established the validity of the reduced-parameter set, we performed the remaining experiments on the reduced-parameter data set only. This is a significant step, since it allows us to solely depend on the 13 parameters obtained from the component loadings, rather than on the PC's from the original 160 values. Furthermore, the results of the first two steps indicate that the RBF network ensembles not only duplicate and surpass the MSA results, but also outperform the MLP algorithms at every step. Since the MLP ensembles fail to show any areas where they may be more desirable than the RBF network ensembles, we will restrict the remainder of the experiments to RBF network ensembles only.

In Sections IV-A and IV-B, we used the two constituent algorithms separately, to discriminate different types of normal tissue from SIL's. In order to obtain the final discrimination between normal tissue and SIL's, *constituent* algorithms (1) and (2) need to be used sequentially. This two-step approach, highlighted in Fig. 4, was specifically developed for the multivariate statistical analysis, which performed best when the decision tasks were simplified. In the next section we present a more direct approach that uses the strengths of the

TABLE VI
ONE-STEP RBF ALGORITHM COMPARED TO MULTISTEP
MSA. (BASED ON REDUCED-PARAMETER SET)

Algorithm	Specificity	Sensitivity
2-step MSA	65%	84%
2-step RBF-ave	65% \pm 2%	87% \pm 1%
2-step RBF-med	67% \pm 2%	87% \pm 1%
1-step RBF-single	65% \pm 9%	89% \pm 2.8%
1-step RBF-ave	67% \pm .7%	91% \pm 1.5%
1-step RBF-med	65.5% \pm .5%	91% \pm 1%

neural network ensembles to reduce the multistep classification scheme to a direct, one-step, process.

C. Single-Step Classification Using Reduced-Parameter Set

In this section, we examine the potential of RBF ensembles for separating SIL's from normal tissue samples in a single classification step. Because the preprocessing for algorithms (1) and (2) is different,¹⁰ we formed 26-dimensional inputs by concatenating the two reduced features sets describes in Section III-A2. We initialized ten kernels using a k -means algorithm on a trimmed¹¹ version of the training set. Without this trimming process, few patterns belonging to the columnar normal class are selected as kernels due to their low prominence in the training set, resulting in poor performance when such a sample is seen. During training, the kernel locations and spreads were not adjusted, to allow kernels to remain in sections of the input space where few patterns are observed. The cost of a misclassified SIL was set at 2.5 times the cost of a misclassified normal tissue sample, in order to provide the best sensitivity/specificity pair. The average and median combiner results are obtained by pooling 20 RBF networks.¹²

Table VI shows the performance of the algorithms for a given SIL misclassification cost. For comparison purposes, the results of the two-step RBF ensemble algorithms are also provided. These algorithms perform the same steps as the MSA, using the results presented in Section III-A2 for each constituent algorithm).

As we discussed above, for an application such as precancer detection, it is far more critical to increase the classification accuracy of some classes than others, to eliminate certain types of errors. By making the algorithm more compact, the one-step algorithm also makes this tradeoff more visible.

1) *Sensitivity-Specificity Tradeoff*: In this subsection we detail the interaction between the cost of misclassification and the variation in the sensitivity/specificity of the RBF ensembles. Since it may be required to reach a predetermined sensitivity, we have varied the cost of misclassifying a SIL to obtain a wide range of sensitivity/specificity pairs. Table VII shows the specificity and sensitivity for various costs of a misclassification.

¹⁰Normalization versus normalization followed by mean scaling.

¹¹The trimmed set has the same number of patterns from each class. Thus, it forces each class to have a similar number of kernels. This set is used *only* for initializing the kernels, *not* while training.

¹²This procedure is repeated 10 times, in order to determine the variability in the ensembles.

TABLE VII
EFFECT OF MISCLASSIFICATION COST ON ONE-STEP RBF ALGORITHM

Algorithm	Cost of SIL misclassification	Specificity	Sensitivity
2-step MSA		65%	84%
RBF-ave	1	85%	61%
RBF-med		84%	61%
RBF-ave	2	75%	88%
RBF-med		74%	86%
RBF-ave	2.5	67%	91%
RBF-med		66%	91%
RBF-ave	3	63%	93%
RBF-med		59%	93%
RBF-ave	4	55%	95%
RBF-med		52%	97%
RBF-ave	5	39%	97%
RBF-med		37%	97%

TABLE VIII
ONE-STEP RBF ALGORITHM COMPARED TO MULTISTEP MSA [18] AND CLINICAL
METHODS [5] FOR DIFFERENTIATING SIL'S AND NORMAL TISSUE SAMPLES

Algorithm	Specificity	Sensitivity
2-step MSA	65%	84%
RBF-single	65% \pm 9%	89% \pm 2.8%
RBF-ave	67% \pm .75%	91% \pm 1.5%
RBF-med	65.5% \pm .5%	91% \pm 1%
Pap smear (human expert)	68% \pm 21%	62% \pm 23%
Colposcopy (human expert)	48% \pm 23 %	94% \pm 6%

On observing the performance of the RBF ensembles at various costs of misclassifying SIL's, the improvements over the two-step MSA algorithm are apparent. If the specificity is required to be above 60%, for example, using a SIL misclassification cost of three improves the sensitivity of the MSA by a significant 10%, using the average RBF ensembles. If, on the other hand, a sensitivity above 83% is required, using a SIL misclassification cost of two, provides an improvement of 12% over the specificity of the MSA.

D. Comparative Results

In the previous sections we discussed how the neural network ensembles were applied to various stages of this classification tasks. In this section, we compare the final results of the RBF network ensembles to the best MSA result and to clinical methods. The SIL misclassification cost of 2.5 provided the best compromise between sensitivity and specificity. To find the variability of the methods, we performed the ensemble averaging ten times on 20 different individual classifiers.

The results of both the two-step and one-step RBF algorithms and the results of the two-step MSA are compared to the accuracy of Pap-smear screening and colposcopy in expert hands in Table VIII. A comparison of one-step RBF algorithms to the two-step RBF algorithms indicates that the one-step algorithms have similar specificities, but a moderate improvement in sensitivity relative to the two-step algorithms. Compared to the MSA, the one-step RBF algorithms have

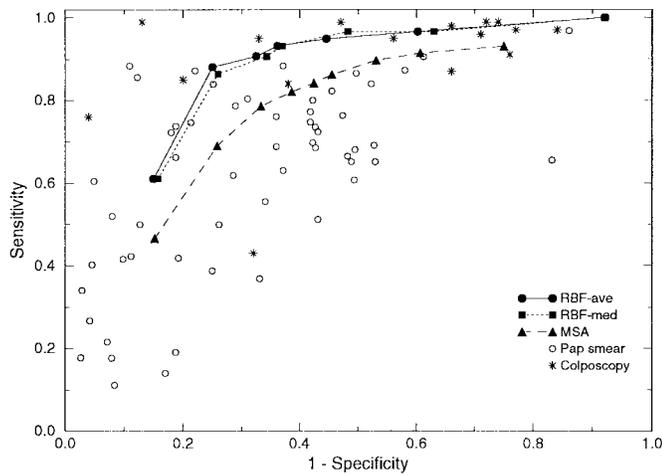


Fig. 5. Tradeoff between sensitivity and specificity for MSA [18] and RBF ensembles. For reference, Pap smear and colposcopy results from the literature on various data sets are included [5].

a slightly decreased specificity, but a substantially improved sensitivity. In addition to the improved sensitivity, the one-step RBF algorithms simplify the decision making process. A comparison between the one-step RBF algorithms and Pap-smear screening indicates that the RBF algorithms have a nearly 30% improvement in sensitivity with no compromise in specificity; when compared to colposcopy in expert hands [5], the RBF ensemble algorithms maintain the sensitivity of expert colposcopists, while improving the specificity by almost 20%.

Fig. 5 further illustrates the tradeoff between specificity and sensitivity for clinical methods, MSA and RBF ensembles, obtained by changing the misclassification cost. The one-step RBF ensembles provide better sensitivity and higher reliability than any other method for a given specificity value.

V. DISCUSSION

In this paper, we demonstrate that cervical tissue fluorescence spectra can be used to develop detection algorithms that differentiate SIL's from normal tissue samples. Of the various algorithms explored, the RBF network ensemble proved to be the best alternative, surpassing single networks, MLP ensembles, and the MSA.

The classification results of both the MSA and the RBF network ensembles demonstrate that significant improvement in classification accuracy can be achieved over current clinical detection modalities using cervical tissue spectral data obtained from *in vivo* fluorescence spectroscopy. The one-step RBF algorithm has the potential to significantly reduce the number of precancerous cases missed by Pap-smear screening and the number of normal tissues misdiagnosed by expert colposcopists.

The qualitative nature of current clinical detection modalities leads to a significant variability in classification accuracy. For example, estimates of the sensitivity and specificity of Pap-smear screening have been shown to range from 11%–99% and 14%–97%, respectively [5]. This limitation can be addressed by exploiting the variance reducing properties of an ensemble approach. In particular, RBF ensembles demonstrate a signifi-

cantly smaller variability in classification accuracy, therefore, enabling more reliable classification. In addition to demonstrating a superior sensitivity, the RBF ensembles simplify the decision-making process of the two-step algorithms based on RBF and MSA into a single step that discriminates between SIL's and normal tissues. We note that for the given data set, both MSA and MLP were unable to provide satisfactory solutions in one step.

The one-step algorithm development process can be readily implemented in software, enabling automated detection of cervical precancer. It can potentially provide near real-time implementation of precancer detection in the hands of nonexperts, and could lead to wide-scale implementation of screening and diagnosis, and more effective patient management in the prevention of cervical cancer. The success of this application will represent an important step forward in both medical laser spectroscopy and gynecologic oncology.

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Kagan Tumer (S'93–M'96) received the B.S. degree in electrical engineering with distinction from George Mason University, Fairfax, VA, in 1989. He received the M.S. degree in engineering from the University of Texas at Austin in August 1992. He completed the Ph.D. degree in information systems engineering at The University of Texas at Austin, Austin, in 1996.

He worked at the Austin Research Center from 1989 to 1991 and at Tracor Applied Sciences during the Summer of 1992. He was a Visiting Professor at

the University of Texas at San Antonio in the 1996/1997 academic year, where he taught information theory and neural networks for pattern recognition, along with introductory courses. He is currently a Computer Scientist at NASA Ames Research Center/Caelum Research and is working on artificial collective intelligences. His research interests include generalization properties of neural networks, multiple classifier combining strategies, and the emerging properties of large complex systems.

Dr. Tumer is a member of the IEEE Computer society, ACM, and honor societies Alpha Chi and Eta Kappa Nu.

Nimmi Ramanujam received the B.S. degree in mechanical engineering from the University of Texas at Austin in 1989. She received the M.S. degree in mechanical engineering and the Ph.D. degree in biomedical engineering, in 1992 and 1995, respectively also from the University of Texas.

In 1996, she joined the Biochemistry and Biophysics Department at the Johnson Foundation at the University of Pennsylvania, Philadelphia, as a Post-Doctoral Fellow. Her research focuses on the application of optical spectroscopy for characterization and detection of disease processes in tissue. Her current research interests include the study of tissue metabolism and oxygenation for characterization of tumor biology using optical methods and the development and application of near infrared photon migration techniques for monitoring and imaging blood oxygenation and hemodynamics *in situ*.

Dr. Ramanujam received the National Institutes of Health Post-Doctoral Fellowship in 1997 for research on tissue optical spectroscopy. She is a member of the Optical Society of America and is currently serving as the Vice-Chair of its Biomedical Optical Spectroscopy Technical Group. She is also a member of the American Society for Laser Medicine and Surgery and the New York Academy of Sciences.



Joydeep Ghosh received the B.Tech degree in 1983 from Indian Institute of Technology (IIT), Kanpur, India, and the M.S. and Ph.D. degrees in 1988 from the University of Southern California, Los Angeles.

He is currently an Associate Professor with the Department of Electrical and Computer Engineering at the University of Texas at Austin, where he holds the Endowed Engineering Foundation Fellowship. He directs the Laboratory for Artificial Neural Systems (LANS), where his research group is studying

the theory and applications of adaptive pattern recognition and multilearner systems. He has published more than 120 refereed papers and edited six books. He is an Associate Editor of *Pattern Recognition*, *Neural Computing Surveys* and *The International Journal of Smart Engineering Design*

Dr. Ghosh received the 1992 Darlington Award for the Best Paper in the areas of CAS/CAD, in addition to "best conference paper" citations for five neural network papers. He served as the General Chairman for the SPIE/SPSE Conference on Image Processing Architectures, Santa Clara, Feb. 1990, as Conference Co-Chair of Artificial Neural Networks in Engineering (ANNIE) from 1993 through 1996, and on the program committee of several conferences on neural networks and parallel processing. He is an Associate Editor and Letters Editor of IEEE TRANSACTIONS ON NEURAL NETWORKS. He was a plenary speaker for ANNIE'97.



Rebecca Richards-Kortum received the B.S. degree in physics and mathematics with highest distinction from the University of Nebraska at Lincoln in 1985. She received the M.S. and Ph.D. degrees in medical physics from the Massachusetts Institute of Technology in 1987 and 1990.

In 1990, she joined the faculty of the Department of Electrical and Computer Engineering and the Biomedical Engineering Program at the University of Texas at Austin. Her research focuses on the application of optical spectroscopy for detection of precancer and other diseases. Her research goals range from describing the basic physics of light propagation in scattering tissues to developing hardware and software systems for the collection and analysis of optical images in the clinical setting. She is a Topical Editor for *Applied Optics* and *The Journal of Biomedical Optics*.

Dr. Richards-Kortum is the 1991 recipient of the National Science Foundation Presidential Young Investigator Award for research in optical spectroscopy. In 1992, she received the Presidential Faculty Fellow Award from the National Science Foundation for excellence in research and teaching.