Objective Detection and Delineation of Oral Neoplasia Using
Autofluorescence Imaging

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Full Text Cancer Prevention Research

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Abstract:

Although the oral cavity is easily accessible to inspection, patients with oral cancer most often present at a late stage, leading to high morbidity and mortality. Autofluorescence imaging has emerged as a promising technology to aid clinicians in screening for oral neoplasia and as an aid to resection, but current approaches rely on subjective interpretation. We present a new method to objectively delineate neoplastic oral mucosa using autofluorescence imaging.

Autofluorescence images were obtained from 56 patients with oral lesions and 11 normal volunteers. From these images, 276 measurements from 159 unique regions of interest (ROI) sites corresponding to normal and confirmed neoplastic areas were identified. Data from ROIs in the first 46 subjects was used to develop a simple classification algorithm based on the ratio of red-to-green fluorescence; performance of this algorithm was then validated using data from the ROIs in the last 21 subjects. This algorithm was applied to patient images to create visual disease-probability maps across the field of view. Histologic sections of resected tissue were used to validate the disease-probability maps.

The best discrimination between neoplastic and non-neoplastic areas was obtained at 405 nm excitation; normal tissue could be discriminated from dysplasia and invasive cancer with a 95.9% sensitivity and 96.2% specificity in the training set and with a 100% sensitivity and 91.4% specificity in the validation set. Disease probability maps qualitatively agreed with both clinical impression and histology.

Autofluorescence imaging coupled with objective image analysis provided a sensitive and non-invasive tool for the detection oral neoplasia.
Introduction

Head and neck cancer, including cancers of the oral cavity, currently ranks as the sixth most common malignancy in the world. There were more than 270,000 new cases of oral cancer reported in 2002. Approximately 60% of these individuals present with stage III or IV disease, and about half will die within five years of diagnosis. Screening individuals at risk for oral cancer and its precursors has the potential to improve early detection, providing the opportunity to intervene when treatment is most effective. In addition, surveillance of patients who have survived their initial oral cancer is important to identify local recurrences and second primary oral tumors, which occur at a higher rate than for any other tumor.

Conventional oral examination using incandescent white light is the current standard of care for screening and surveillance for oral cancer and precancerous lesions. The sensitivity of visual examination is limited by several factors including the experience and index of suspicion of the examiners. In primary care situations, cases of malignancy may be seen rarely and clinicians may have difficulty discriminating the sometimes subtle mucosal changes associated with premalignant lesions and early cancers from more common benign inflammatory conditions. Furthermore, it can be challenging to delineate the boundaries of neoplastic lesions using conventional oral examination making the choice of a biopsy location difficult.

Several new approaches have been proposed to address the limitations of the conventional oral examination, including the use of toluidine blue, brush cytology,
reflectance visualization after acetic acid application, and illumination with a chemiluminescent light source. While useful in certain situations, each of these approaches is associated with a high rate of false-positives 5-8. Recently, several studies have demonstrated that autofluorescence imaging may improve the ability to distinguish normal from premalignant and malignant oral tissue 9-15. When tissue is illuminated in the ultraviolet-visible region, a portion of photons are absorbed by molecules within the tissue called fluorophores which then emit lower energy photons that can be detected as fluorescence from the mucosal surface. Examples of fluorophores which produce autofluorescence signals in tissue include NADH and FAD in the epithelial layer, and collagen and elastin crosslinks in the stroma 16. In comparison to normal oral tissue, neoplastic lesions are associated with a decrease of green fluorescence when excited with ultraviolet (UV) or near-UV light 9,10,12,17 that is attributed to decreased signal from collagen crosslinks in the stroma 18. Increased red fluorescence has also been observed by several groups in oral lesions and is frequently attributed to porphyrins 19. Several groups have proposed that this perceived loss of green fluorescence and increase in red fluorescence can be useful as a diagnostic aid to help detect and diagnose early neoplastic disease in several anatomic sites including the oral cavity, bronchus, cervix, esophagus and colon 13,20-23. In addition, the changes in fluorescence may aid in surgical resection by delineating the extent of neoplastic changes beyond the clinically apparent margins 9,10.

Recently, the U.S. Food and Drug Administration approved an autofluorescence imaging device for early detection of oral neoplasia. The device, marketed as the VELscope® (LED Dental, Inc., White Rock, BC, Canada), uses a blue/violet light (400 –
460 nm wavelengths) to illuminate oral tissue and long pass and notch filters to enable clinicians to directly visualize fluorescence in the oral cavity\(^9,13\). The VELscope and other proposed fluorescence imaging devices rely on qualitative observations to detect and delineate neoplastic oral lesions and therefore reliable screening with these instruments necessitates well-defined and standardized image interpretation criteria, and appropriate user training. This may not be feasible in many primary care situations. We hypothesize that the application of digital image processing techniques to autofluorescence imaging of oral tissue will provide the ability to objectively identify and delineate the peripheral extent of neoplastic lesions in the oral cavity. This will provide a powerful tool in patient care locations where experts are not available or where physicians encounter few cases of malignant and premalignant neoplasia. Low-cost digital cameras with sufficient sensitivity to record tissue autofluorescence in near real time are now readily available\(^{24}\), making clinical application of such automated image processing feasible.

The primary goal of the present study was to evaluate the use of quantitative autofluorescence imaging for the detection and delineation of oral neoplastic lesions. We demonstrate that a simple, objective method can be used to accurately classify regions of interest within an autofluorescence image with 100\% sensitivity and 91.4\% specificity relative to histopathology. This method can delineate the presence and extent of neoplastic lesions within a field of view and provide results which correlate with the histopathologic assessment of extent of disease. Thus, quantitative autofluorescence
imaging may provide a non-invasive and objective method to improve screening and margin delineation of oral cancers and precancers.

Methods

Human Subjects

Study subjects were enrolled in a clinical protocol reviewed and approved by the Internal Review Boards at The University of Texas MD Anderson Cancer Center and Rice University. Patients were eligible and recruited if they were 18 years of age or older and had known or suspected precancerous or cancerous squamous lesions located in the oral mucosa. Patients may have had previous surgical, radiation, or chemotherapeutic treatments. Normal volunteers were eligible and recruited if they were 18 years of age or older and had no history of oral pathology. All subjects enrolled in the study gave written informed consent. The average age of patients in this study was 59, 42% of the patients were female and 58% were male. The average age of normal volunteers in this study was 27.4, 27% were female and 73% were male.

Imaging Procedure

Autofluorescence images were obtained from the oral cavity of 56 patients with clinically abnormal lesions and 11 normal volunteers. Data were divided into a training set and a validation set. Data acquired from the first 39 patients and 7 normal volunteers imaged between June 2006 and January 2008 were allocated to the training set and used
to develop an algorithm for detection of neoplasia. Data acquired from the subsequent 17 patients and 4 normal volunteers imaged between March and June 2008 formed a validation set and were used to test the performance of this algorithm relative to histopathology.

White light and autofluorescence images were obtained at 365 nm, 380 nm, 405 nm, and 450 nm excitation using a Multispectral Digital Microscope (MDM). This device is described in detail elsewhere but briefly, the MDM is a wide-field optical microscope which collects digital autofluorescence and reflectance images with a color CCD camera from a variable field of view, ranging in size from approximately 1 to 7 cm. Patients were imaged either in an outpatient clinic or in the operating room under general anesthesia prior to surgery. A physician positioned the patient and microscope so that the suspicious lesion or area of interest was clearly in the field of view of the device. Clinically normal areas distant from or contralateral to the lesion were also imaged. Following imaging in the clinic, suspicious lesions were biopsied. In the operating room, previously biopsied lesions were surgically resected.

**Histopathologic Correlation**

Biopsies and resected tissues were evaluated using standard histopathologic analysis by a board certified pathologist (either AEN or MDW). The location of biopsies and resected lesions was recorded using digital photography so that pathology results could later be correlated to multispectral imaging results. In addition, the locations of gross anatomical features were noted in both autofluorescence images and
histology specimens to aid in correlation. The resulting histopathology sections were evaluated to provide a diagnosis along the entire length of the epithelium, also noting any submucosal abnormalities in each slide. Histopathology diagnosis included the following categories: normal, mild dysplasia, moderate dysplasia, severe dysplasia/carcinoma in situ, and invasive carcinoma. For the purposes of diagnostic algorithm development, two major categories were defined: normal tissue (including inflammation and hyperplasia) and neoplastic tissue (including dysplasia, carcinoma in situ and cancer).

**Analysis and Statistical methods**

Images were preprocessed to subtract signal from ambient room light and translated so that white light and fluorescence images of the same field of view were spatially registered. 276 measurements corresponding to 159 unique regions of interest (ROIs) sites of clinically normal and suspicious regions of tissue were selected from white light images by a head and neck surgeon (AMG) blinded to the results of the autofluorescence imaging. In some cases, repeat measurements were obtained from the same ROI site to help ensure image data was collected without motion artifacts; often both the first and repeat measurements were included in the analysis. These repeat measurements account for the difference between the number of measurements and the number of ROI sites. Heterogeneity in pathologic diagnoses may occur within relatively small areas of diseased oral mucosa\(^{26,27}\) so ROIs were stringently selected from suspicious areas using one of following four criteria: 1) areas corresponding to the same size and location as a biopsy with a pathological diagnosis, 2) ROIs from locations
which could be correlated to a histopathology slide with a corresponding pathological
diagnosis, 3) areas within well-defined exophytic tumors confirmed by pathological
diagnosis and 4) ROIs from a location which was clinically normal and deemed by the
physician to be sufficiently distant from the lesion.

Autofluorescence images from the training set were analyzed to determine
whether specific image features could be used to classify a measurement site as normal or
neoplastic. The autofluorescence images and white light images were spatially registered
so that the ROIs chosen in the white light images corresponded to the same region of
tissue in the autofluorescence images. The training set included data from the first 39
patients and 7 normal volunteers and included measurements from 173 measurements
from 102 unique ROIs. Qualitatively, neoplastic ROIs were associated with a decrease in
average green fluorescence intensity and often an increase in red fluorescence intensity.
The mean ratio of red-to-green pixel intensities inside each of the ROIs was calculated
from the fluorescence images at each excitation wavelength. Red and green pixel
intensities were obtained from the collected Red-Green-Blue color images, created by the
Bayer color mask on the CCD detector. A classifier was developed to distinguish
neoplastic and normal ROIs using linear discriminant analysis with the single input
feature of average ratio of red-to-green fluorescence. When more than one measurement
corresponded to a ROI site, the mean of the feature values was used for classification.
The classifier was trained using all of the ROI sites in training set and the prior
probability input into the classifier was chosen to represent the percentage of abnormal to
normal measurements in the data set. The classifier was developed after images were
acquired from patients in the training set but before measurements were acquired from patients in the validation set. Classifier accuracy in the training set was assessed by plotting the receiver operating characteristic (ROC) curve, the area under the ROC curve (AUC), and the sensitivity and specificity at a particular operating point on the ROC curve. The positive and negative predictive values were also calculated at the operating point. Confidence intervals were calculated for operating characteristics using the Wilson ‘score’ method including a continuity correction.

The algorithm was then applied to data from the validation set using the red-to-green ratio threshold found to produce the highest combination of sensitivity and specificity in the training set. The validation set was designed to rigorously test the algorithm and for most patients, ROI and biopsy pairs were collected on the clinical margins of the lesion in addition to directly on the lesion and in clinically normal areas. The validation set included 103 measurements from 57 unique ROIs in a second group of 17 patients and 4 normal volunteers.

An additional analysis step was explored to increase the performance of the classifier by normalizing the red-to-green ratio measurements for each patient. An additional unique and non-overlapping ROI of clinically normal tissue was chosen from the same anatomical site and in the same field of view for each of the ROIs described above. At each excitation wavelength, the mean red-to-green autofluorescence ratio was calculated in this ROI; the mean red-to-green ratios from the other ROIs were normalized by this value. This method provides a way to compensate for anatomical and patient to
patient variations in red-to-green fluorescence intensity ratio. Identical statistical analysis was performed using this measured feature with both the training set and the validation set. The method utilizing the magnitude of the red-to-green fluorescence intensity ratio is termed the *raw red-to-green method* and the method utilizing a normalized red-to-green fluorescence intensity ratio is termed *normalized red-to-green method*.

**Disease Probability Maps**

The classification algorithms described above provided a relationship between the magnitude of the red-to-green fluorescence intensity ratio for a particular region of interest within the image and the probability of that region having a diagnosis of abnormal. This relationship was used to predict the probability of a diagnosis of dysplasia or cancer for each pixel in an image, given the red-to-green fluorescence intensity ratio at that pixel. The posterior probability values at each pixel in the image were computed and pixels which corresponded to a 50% or greater probability of being classified as dysplastic or cancerous were color coded and digitally overlaid onto the white light images. This method provides a means to illustrate areas of tissue with the highest probability of being neoplastic. The assumption was made that the region of interest method described above could be generalized on a pixel by pixel basis. Disease probability maps were compared to histologic images of tissue resected from the field of view to confirm the accuracy of this method.
Results

Tables 1 and 2 summarize the anatomic site and histopathologic diagnoses of the 159 sites included in this analysis. The most common sites were tongue, buccal mucosa and floor of mouth, followed by palate, lip, and gingiva. The training set contained 52% normal, 28% dysplastic, and 20% invasive carcinoma sites while the validation set contained 61% normal, 26% dysplastic, and 12% invasive carcinoma sites. The normal histopathologic category could include tissue with hyperkeratosis, hyperplasia, and/or inflammation as long as there was no dysplasia or carcinoma. The normal sites in the training set, based on available pathology (not including normal volunteers and normal sites where no biopsy was taken), included 7 sites (13.2% of normal sites) with hyperplasia and hyperkeratosis, 4 sites (7.5% of normal sites) with hyperkeratosis, and 3 sites (5.7% of normal sites) with hyperplasia and/or fibroadipose tissue. The validation set included 3 sites (8.6% of normal sites) with hyperplasia and hyperkeratosis, 1 sites (2.9% of normal sites) with hyperplasia, 1 site (2.9% of normal sites) with a submucosal hemorrhage, and 1 site (2.9% of normal sites) with marked inflammation and osteonecrosis. The abnormal histopathology category could include dysplasia and carcinoma. In the training set 59.2% of the abnormal sites were premalignant (mild, moderate, or severe dysplasia), in the validation set 68.2% of the abnormal sites were premalignant.

Figure 1 shows white light and autofluorescence images from the buccal mucosa of a patient with pathologically confirmed invasive carcinoma. The white light image
(Fig. 1A) shows two ROIs, one which corresponds to a pathologically confirmed invasive carcinoma, and the other which was clinically normal and outside of the pathologically confirmed clear resection margin. Figures 1B-1D show autofluorescence images at different excitation wavelengths that were taken before surgery from the same field of view. The autofluorescence image obtained at 405 nm excitation qualitatively shows the greatest visual contrast between the normal and neoplastic ROI. This observation was typical for study patients.

Table 3 summarizes the performance of both diagnostic algorithms, based on either the raw or the normalized mean red to green fluorescence intensity ratios, for classifying lesions in the training set. At each excitation wavelength, the classifier that used the normalized red-to-green fluorescence intensity ratio (Normalized R/G ratio) had slightly higher AUC than the algorithm based on the raw red/green fluorescence intensity ratio (Raw R/G ratio). In all cases, the highest AUC was obtained at 405 nm excitation. The sensitivity and specificity values at the point on the ROC curve nearest the gold standard (Q-point) are also reported in Table 3.

A scatter plot of the normalized red-to-green ratio at 405 nm excitation for each of the 102 sites in the training set, as well as the threshold of 1.19 used in the classification algorithm is shown in Figure 2A. Of the 102 sites, 4 were misclassified including one site of fibroadipose tissue on the lower lip misclassified at abnormal, one hyperkeratotic site on the right buccal misclassified at abnormal, one cancer site on the right lateral tongue misclassified as normal, and one site on the left soft palate with focal ulceration
and dysplasia misclassified as normal. Figure 2B shows the ROC curve for this classifier; the AUC is 0.988, and at the Q-point, the sensitivity is 95.9% (95% confidence interval (CI) 84.9% - 99.3%) and the specificity is 96.2% (95% CI 85.9% - 99.3%). The positive predictive value is 95.9% (95% CI 84.9% - 99.3%) and the negative predictive value is 96.2% (95% CI 85.9% - 99.3%). This operating point is indicated on the ROC curve.

The algorithm using the normalized red-to-green fluorescence intensity ratio at 405 nm excitation was applied to the validation set. In Figure 2C a scatterplot of the normalized R/G ratio for each site in the validation set is shown along with the threshold that had been previously selected for the training set. Figure 2D depicts the ROC curve with the operating point selected for the training set indicated. A 100% sensitivity (95% CI 81.5% - 99.6%) and 91.4% specificity (95% CI 75.8% - 97.8%) and an AUC of .987 were achieved at this operating point for the validation set. The positive predictive value is 88.0% (95% CI 67.7% - 96.9%) and the negative predictive value is 100% (95% CI 86.7% - 99.7%). Of the 57 sites in the validation set, 3 were misclassified as abnormal including one site on the left buccal with hyperplasia, one site on the right buccal, and another site on the left buccal.

Figure 3 shows white light and 405 nm excited autofluorescence images from a study patient with moderate dysplasia and carcinoma in situ located in the floor of mouth. The white light image is also shown with an overlay of the calculated disease probability map; regions corresponding to a predictive probability of a neoplastic lesion greater than 50% are shaded as indicated by the color bar. The disease probability map indicates the
probability that a particular pixel in the image corresponds to a neoplastic area of tissue. Histologic sections obtained at six areas in the tissue are also shown. Only one of these areas was included in the previous classification analysis. The disease probability map shows qualitative agreement with the presence of dysplasia and cancer in the areas corresponding to the histologic sections.

Figure 4 shows representative white light images with and without superimposed disease probability maps from four study patients. Images in the first three rows correspond to patients with histologically confirmed neoplasia, while the image in the bottom row is from a normal volunteer with no clinically suspicious lesions. Although the lesion in Figure A is obvious, those in Figures B and C are less so, highlighting the potential to aid clinicians in identifying the presence of neoplasia and identifying optimal sites for further evaluation with biopsy. Images in Figures 4A and B are from a patient with an invasive carcinoma in the floor of mouth. Images in Figures 4C and D are from a patient with a region of severe dysplasia on the tongue. The images in Figures 4E and F are from a patient with a region of moderate dysplasia on the gingiva. In all three cases, the disease probability map delineates the suspicious regions identified clinically by an oral cancer specialist blinded to the results of the autofluorescence imaging and are consistent with histopathologic sections obtained. Figures 4G and H are from the inner lip of a normal volunteer and the disease probability map does not indicate any lesions.
Discussion

Our results illustrate how autofluorescence imaging may enhance the ability of clinicians to detect and delineate areas of oral dysplasia and carcinoma. Although all four illumination conditions tested allowed visualization of changes in autofluorescence with neoplasia, illumination with 405 nm wavelength produced the highest discriminatory capability. This corresponds to previous findings comparing illumination wavelengths for autofluorescence imaging in freshly resected oral cancer surgical specimens. While subjective interpretation of loss of autofluorescence has been shown to be useful, there are several important advantages associated with objective and quantitative analysis of changes in autofluorescence signal. First, quantitative analysis methods provide a rigorous and repeatable way to determine the threshold for demarcating a lesion, even for providers with less experience. Second, digital imaging allows the operator to save and process images, directly comparing data from multiple patients in a series or from a single patient over time. Third, ratios of fluorescence intensity values provide a way to reduce variations in images associated with spatial non-uniformities in illumination.

In the present study, the performance of a simple classifier based on the ratio of red-to-green autofluorescence intensity at 405 nm excitation was tested and found to discriminate neoplastic and non-neoplastic tissue with a sensitivity and specificity of 96% in the training set and 100% sensitivity and 91.4% specificity in the validation set. These results compare favorably with the performance of visual oral examination, which has been systematically reviewed by Downer et al. Downer identified eight prospective studies between 1980 and 2002 that involved conventional oral exam with gold standard
verification provided by an expert observer. In four of the studies the screeners were general dentists and in four of the studies the screeners were trained health workers. Sensitivity ranged from 59% to 97%, specificity ranged from 75% to 99%, and meta-analysis resulted in a weighted pooled sensitivity of 85% and a specificity of 97%. Other reports of the performance of visual oral screening include Sankaranarayanan et al (sensitivity 77%, specificity 76%) 33, Ramadas et al (sensitivity 82%, specificity 85%) 34, and Nagao et al (sensitivity 92%, specificity 64%) 35. The classifier in this study can be applied to entire images of the oral cavity to visualize areas with a high probability of being neoplastic; disease probability maps are consistent with histologic sections obtained from tissue in the field of view.

Autofluorescence imaging has shown great promise for enhancing visualization of neoplastic areas in recent studies 9,10,13-15,17. In a study of 44 patients, Lane et al. demonstrated high sensitivity and specificity at discriminating normal oral mucosa from severe dysplasia, carcinoma in situ, or invasive carcinoma based on visual assessment of loss of autofluorescence in diseased mucosa at excitation wavelengths between 400 nm and 460 nm 13. In another study by the same group, the potential for autofluorescence imaging to enhance delineation of the margins of neoplastic changes was demonstrated. In some cases fluorescence loss extended as far as 25 mm beyond the clinically apparent margin 9. Autofluorescence endoscopic imaging technologies for lung and the GI tract utilizing ratios of red and green signal have been available for over a decade and have greatly increased sensitivity of disease detection in these organ systems 20,36-38. The LIFETM (laser induced fluorescence emission) system is an autofluorescence
bronchoscopy device which provides the user with a real time image in which changes in hue correspond to suspicious and/or abnormal areas. Users of the device must be trained in order to interpret these changes in image hue.

A potential confounding factor which may limit specificity of classifiers based on the red-to-green fluorescence intensity ratio for automated image analysis software is the frequent presence of red fluorescence on normal papillae of the dorsal aspect of the tongue. At 405 nm excitation, increased fluorescence above 600 nm emission has been observed in oral lesions and is thought to originate from porphyrins, although it is uncertain whether the origins of these porphyrins are intrinsic or derived from bacterial contamination. Red porphyrin-like fluorescence has been shown to correlate with neoplastic disease in other regions of the oral cavity. However, normal red fluorescence is limited to the dorsal tongue; it is not observed on normal areas on the lateral and ventral tongue where there is a much higher propensity for developing neoplastic disease. In this study, a single site in the training set was imaged on the dorsal tongue and corresponded to cancer. Bright red fluorescence was visible at this site.

Our results demonstrate the potential of quantitative fluorescence imaging as an objective approach to non-invasively identify and delineate the mucosal extent of neoplastic lesions in the oral cavity. It should be noted that the images were obtained with a research-grade device and the disease probability maps described here were constructed subsequent to the image acquisition and compared to only a limited number of sites with diagnosis confirmed with histopathology. Unfortunately, it is difficult to
obtain pathology diagnosis for the entire surface of a resected specimen. Additionally, in order to provide optimal benefit to clinicians both for detection and margin delineation of oral dysplasia and carcinoma, these disease probability maps need to be available to view in real or near real-time. We are currently making software improvements and developing quantitative fluorescence imaging devices that can show false color disease-probability maps based on red/green fluorescence intensity ratios at 405 nm excitation at the time of the examination. In addition, we recognize that our results were obtained from a small group of subjects with disease prevalence that might be expected in a treatment population rather than a screening population. Future studies are planned to evaluate this method of quantitative fluorescence imaging in community settings in a larger subject group with a wide spectrum of oral pathologies including oral dysplasia and early invasive disease, benign conditions and inflammation.

In summary, the present study provides proof-of-principle for the use of a practical tool for the quantitative fluorescence imaging as an objective technique for detection and delineation of oral neoplasia. The use of objective disease probability maps represents an important advance toward integration of optical imaging technologies into the clinical practice of dentists and primary health care workers. Development of non-invasive and objective diagnostic aids based on these findings may facilitate early detection and diagnosis of oral cancer and its precursors by less experienced personnel at the point of care.
References:


Tables

**Table 1.** Anatomic sites of ROIs in the training and validation set. Note: Percentages may not add up to 100 % because of rounding.

<table>
<thead>
<tr>
<th>Anatomical Site</th>
<th>No. of sites in training set (%)</th>
<th>No. of sites in validation set (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Tongue</td>
<td>37 (36.3)</td>
<td>19 (33.3)</td>
</tr>
<tr>
<td>Buccal mucosa</td>
<td>12 (11.8)</td>
<td>15 (26.3)</td>
</tr>
<tr>
<td>Floor of mouth</td>
<td>22 (21.6)</td>
<td>4 (7.0)</td>
</tr>
<tr>
<td>Gingiva</td>
<td>2 (2.0)</td>
<td>7 (12.3)</td>
</tr>
<tr>
<td>Lip</td>
<td>14 (13.7)</td>
<td>4 (7.0)</td>
</tr>
<tr>
<td>Palate</td>
<td>15 (14.7)</td>
<td>8 (14.0)</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td><strong>102 (100)</strong></td>
<td><strong>57 (100)</strong></td>
</tr>
</tbody>
</table>

**Table 2.** Pathology diagnosis of ROI sites in training and validation set.

<table>
<thead>
<tr>
<th>Diagnosis</th>
<th>No. of sites in training set (%)</th>
<th>No. of sites in validation set (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Normal</td>
<td>53 (52.0)</td>
<td>35 (61.4)</td>
</tr>
<tr>
<td>Mild dysplasia</td>
<td>11 (10.8)</td>
<td>5 (8.8)</td>
</tr>
<tr>
<td>Moderate dysplasia</td>
<td>6 (5.9)</td>
<td>4 (7.0)</td>
</tr>
<tr>
<td>Severe dysplasia/CIS</td>
<td>12 (11.8)</td>
<td>6 (10.5)</td>
</tr>
<tr>
<td>Invasive carcinoma</td>
<td>20 (19.6)</td>
<td>7 (12.3)</td>
</tr>
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<td><strong>Total</strong></td>
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</table>
Table 3. Classification results at each fluorescence excitation wavelength using both the 
*Raw R/G Ratio* method and the *Normalized R/G ratio* method in the training set.

<table>
<thead>
<tr>
<th>Fluorescence excitation wavelength</th>
<th>Raw R/G ratio</th>
<th>Normalized R/G ratio</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>AUC</td>
<td>Sensitivity (%)</td>
</tr>
<tr>
<td>365 nm</td>
<td>.832</td>
<td>83.7</td>
</tr>
<tr>
<td>380 nm</td>
<td>.891</td>
<td>89.8</td>
</tr>
<tr>
<td>405 nm</td>
<td>.971</td>
<td>91.8</td>
</tr>
<tr>
<td>450 nm</td>
<td>.922</td>
<td>81.6</td>
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</tbody>
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**Figure Legends**

**Figure 1.** Autofluorescence and white light images of the buccal mucosa of a typical study patient.  
A. White light image showing regions of interest of histopathologically confirmed normal tissue and invasive carcinoma.  
B. Fluorescence image at 365 nm excitation.  
C. Fluorescence image at 405 nm excitation.  
D. Fluorescence image at 450 nm excitation.

**Figure 2.**  
A. Scatter plot of normalized red-to-green ratios at 405 nm excitation for the 102 ROI sites in the training set. The horizontal line indicates the threshold used to obtain 95.9% sensitivity and 96.2% specificity. Note that 2 additional abnormal data points had a red-to-green fluorescence intensity ratio greater than 3 but are not shown on this plot.  
B. Receiver-operating characteristic (ROC) curve of the classifier based on the normalized red-to-green ratio. The operating point used for classification is indicated by
a dot and arrow. C. Scatter plot of the red-to-green ratio for the 57 sites in the validation set with threshold selected from the training set indicated. Note that 3 additional abnormal data points had a red-to-green fluorescence intensity ratio greater than 3 but are not shown on this plot. D. ROC curve obtained for the validation set. The operating point is indicated and corresponds to the threshold chosen from the training set.

**Figure 3.** A. White light image of floor of mouth with histopathologically confirmed dysplasia and carcinoma in situ. B. 405 nm excitation fluorescence image showing areas with deceased autofluorescence. C. White light image with disease probability map showing the predictive probability of a neoplastic lesion superimposed. Letters indicate specific locations were pathology is known. The key to the right of C. indicates pathology. The histology slides below show tissue sections from these areas. Marking bar at the lower right-hand corner = 1 mm.

**Figure 4.** A. and B. show images from a patient with an invasive carcinoma in the floor of mouth. A. White light image B. White light image with disease probability mapping showing the predictive probability of a neoplastic lesion. C. and D. show images from a patient with a region of severe dysplasia on the tongue. E and F show images from a patient with a region of moderate dysplasia on the gingiva. G. and H. show images from inner lip of a normal volunteer.
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<tr>
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<td>22 (21.6)</td>
<td>4 (7.0)</td>
</tr>
<tr>
<td>Gingiva</td>
<td>2 (2.0)</td>
<td>7 (12.3)</td>
</tr>
<tr>
<td>Lip</td>
<td>14 (13.7)</td>
<td>4 (7.0)</td>
</tr>
<tr>
<td>Palate</td>
<td>15 (14.7)</td>
<td>8 (14.0)</td>
</tr>
<tr>
<td><strong>Total</strong></td>
<td><strong>102 (100)</strong></td>
<td><strong>57 (100)</strong></td>
</tr>
<tr>
<td>Diagnosis</td>
<td>No. of sites in training set (%)</td>
<td>No. of sites in validation set (%)</td>
</tr>
<tr>
<td>-------------------------</td>
<td>----------------------------------</td>
<td>-----------------------------------</td>
</tr>
<tr>
<td>Normal</td>
<td>53 (52.0)</td>
<td>35 (61.4)</td>
</tr>
<tr>
<td>Mild dysplasia</td>
<td>11 (10.8)</td>
<td>5 (8.8)</td>
</tr>
<tr>
<td>Moderate dysplasia</td>
<td>6 (5.9)</td>
<td>4 (7.0)</td>
</tr>
<tr>
<td>Severe dysplasia/CIS</td>
<td>12 (11.8)</td>
<td>6 (10.5)</td>
</tr>
<tr>
<td>Invasive carcinoma</td>
<td>20 (19.6)</td>
<td>7 (12.3)</td>
</tr>
<tr>
<td>Total</td>
<td>102 (100)</td>
<td>57 (100)</td>
</tr>
<tr>
<td>Fluorescence excitation wavelength</td>
<td>Raw R/G ratio</td>
<td>Normalized R/G ratio</td>
</tr>
<tr>
<td>-----------------------------------</td>
<td>--------------</td>
<td>----------------------</td>
</tr>
<tr>
<td></td>
<td>AUC</td>
<td>Sensitivity (%)</td>
</tr>
<tr>
<td>365 nm</td>
<td>.832</td>
<td>83.7</td>
</tr>
<tr>
<td>380 nm</td>
<td>.891</td>
<td>89.8</td>
</tr>
<tr>
<td>405 nm</td>
<td>.971</td>
<td>91.8</td>
</tr>
<tr>
<td>450 nm</td>
<td>.922</td>
<td>81.6</td>
</tr>
</tbody>
</table>
Training Set

A. 405 nm Fluorescence Red/Green Ratio
B. Sensitivity: 95.9%
Specificity: 96.2%
AUC: .988

Validation Set

C. 405 nm Fluorescence Red/Green Ratio
D. Sensitivity: 100%
Specificity: 91.4%
AUC: .987