1 Q1 Research Article

40

41

42

43

44

45

46

47

48

49

50

51

A Pilot Study of Low-Cost, High-Resolution Microendoscopy as a Tool for Identifying Women with Cervical Precancer

5 AU Mark C. Pierce¹, YaoYao Guan², Mary Kate Quinn³, Xun Zhang⁴, Wen-Hua Zhang⁴, You-Lin Qiao⁴, 6 Philip Castle⁵, and Rebecca Richards-Kortum³

Abstract

7

8

9

10

11

12 13

14 15

16

17 18

19

20

21

22

23

24

29

30

31

32

33

34

35

36

37

38

Cervical cancer remains one of the leading causes of death among women in developing countries. Without resources to support Pap smear cytology and colposcopy, cost-effective approaches which enable single-visit "see-and-treat" protocols offer the potential to reduce morbidity and mortality due to this preventable disease. We carried out a pilot clinical study in Shanxi province, China, to evaluate a low-cost, high-resolution microendoscope (HRME) imaging system which enables evaluation of epithelial cell morphology *in vivo*. HRME images were obtained at discrete sites on the cervix in 174 women, in addition to visual inspection with acetic acid (VIA) and colposcopic examination. Of 69 sites appearing abnormal on colposcopy, only 12 showed high-grade disease (CIN2+) on pathology. Quantification of the nuclear-to-cytoplasm ratio by HRME enabled an *ad hoc* threshold to be defined, which correctly classified all 12 sites as abnormal, whilst classifying 38 of the remaining 57 pathology normal sites as normal. All patients with biopsy confirmed high-grade disease also tested positive for high-risk human papilloma virus (HPV) DNA and were classified as abnormal by HRME. Among the remaining patients who tested positive for HPV but were either normal by colposcopy or showed <CIN2 on pathology, only 6 of 32 (18.8%) were classified as abnormal by HRME.

Visual examination techniques for cervical cancer screening may overestimate the prevalence of precancerous lesions, leading to unnecessary treatment, expense, and patient stress. The results of this study suggest that evaluation of suspicious lesions by HRME may assist in ruling out immediate cryotherapy, thus increasing the efficiency of current see-and-treat programs. *Cancer Prev Res; 1–7. 2012 AACR*.

Introduction

Cervical cancer is the third most common cancer amongst women worldwide; an estimated 530,000 new cases occurred and 275,000 women died from this treatable disease in 2008 (1). More than 80% of these cases occur in developing countries (2), which lack the resources and expertise required to maintain the regular screening programs used in industrialized nations. In low-resource settings, techniques such as visual inspection with acetic acid (VIA) or with Lugol's iodine (VILI) have been proposed as

Note: Supplementary data for this article are available at Cancer Prevention Research Online (http://cancerprevres.aacrjournals.org/).

M.C. Pierce and Y.Y. Guan contributed equally to this work.

Corresponding Author: Rebecca Richards-Kortum, Rice University, 6100 Main Street, Houston, TX 77005. Phone: 713-348-3823; Fax: 713-348-5877; E-mail: rkortum@rice.edu

doi: 10.1158/1940-6207.CAPR-12-0221

2012 American Association for Cancer Research.

cost-effective alternatives to traditional Pap/cytology programs for cervical cancer screening. In several large clinical studies, VIA has shown clinical sensitivity ranging from 41% to 92%, approaching that of standard colposcopy (3–5). Such methods have enabled "see-and-treat" programs to be implemented, using cryotherapy for immediate ablation of any lesion appearing abnormal by VIA. The ability to deliver diagnostic and therapeutic services in a single clinic visit is a key factor in reducing patient loss to follow-up after a positive screening test, which can amount to 15% of patients or more when a multi-visit screening approach is required (6).

While the sensitivity of VIA/VILI is quite good, some 52studies have reported specificity figures as low as 49% 53(7). Poor specificity, along with the potential for loss to 54follow-up, has raised concerns that see-and-treat programs 55using VIA/VILI may lead to overtreatment of many benign 56conditions which do not represent significant cervical can-57cer risk and will resolve without intervention. Overtreat-58ment raises the expense of these programs and may cause 59unnecessary concern for the patient. In the absence of 60 colposcopically guided biopsy collection with histopathol-6162ogy processing and review, new approaches are required to identify those patients who genuinely require treatment. 63 Optical imaging and spectroscopy techniques have been 64

Authors' Affiliations: ¹Department of Biomedical Engineering, Rutgers, The State University of New Jersey, Piscataway, New Jersey; ²Johns Hopkins University School of Medicine, Baltimore Maryland; ³Department of Bioengineering, Rice University, Houston, Texas; ⁴Cancer Institute, Chinese Academy of Medical Sciences, Beijing, China; and ⁵American Society for Clinical Pathology, Washington, District of Columbia

67 shown to detect alterations in tissue morphology and bio-68 chemistry within epithelial and stromal tissue components. 69 associated with the onset and progression of cervical neo-70 plasia (8-10). Macroscopic optical imaging (similar to 71standard colposcopy) examines an entire organ surface 72under white light, narrow-band illumination, and/or under 73conditions required for fluorescence excitation. In contrast, 74microscopic optical imaging involves placement of the tip of a small fiber optic probe directly onto the cervical 7576epithelium, enabling individual cells to be visualized in vivo (11). By using exogenous contrast agents such as acetic acid 77 78 or acriflavine/proflavine, morphologic features used by pathologists such as nuclear crowding, pleomorphism, and 7980 nuclear-to-cytoplasm ratio can be assessed in vivo and in real-time (12, 13). 81

82 We describe here the results of a pilot clinical study using 83 a recently developed low-cost microscopic imaging system, 84 termed the high-resolution microendoscope (HRME; 85 ref. 14), for evaluation of cervical lesions appearing abnor-86 mal under VIA or colposcopy. Our long-term hypothesis 87 is that HRME imaging can improve the specificity of early 88 detection of cervical cancer and its precursors by ruling out 89 many of the visually or colposcopically apparent lesions 90 that are actually benign. Here, we set out to establish 91whether HRME can identify cervical lesions which do not 92require treatment in patients initially screened by VIA or 93 human papilloma virus (HPV) testing. Reducing the numbers of lesions treated unnecessarily following visual exam-94 95ination or colposcopy would clearly benefit the patient 96 while also lowering the overall costs of see-and-treat pro-97 grams in the settings where their impact is greatest.

98 Materials and Methods

Study population

Institutional Review Boards at each of the clinical sites and academic institutions involved, including Johns Hopkins University (Baltimore, MD), the Chinese Academy of 103Medical Sciences (Beijing, China), and Rice University 104(Houston, TX), approved the study. A total of 2,500 women 105older than 18 years living in Xiangyuan County, China, 106 underwent initial VIA/VILI examination as part of their 107 involvement in the national cancer screening program. On 108the basis of prior data from the program, it was estimated 109that 4% of this cohort would receive a positive VIA/VILI test 110 result (Fig. 1A). As a nested pilot study to evaluate HRME, 111 112patients with an abnormal VIA/VILI examination (n = 63) were invited to participate in this study, along with a 113 random selection of patients with a normal VIA/VILI exam-114 ination (n = 111). The imaging portion of the study was 115completed in August 2010. The mean, median, interguartile 116range, and total range of age for the 174 subjects were 41, 41, 11736-45, and 29-58 years, respectively. 118

119

120

121

122

123

124

125

126

127

128

129

130

131

132

133

134

135

136

137

Study procedures

The 174 participants were offered transportation from their local villages to the Xiangyuan County Maternal and Child Health Hospital of Shanxi, where the study was conducted (Fig. 1B). Each participant had an initial oneon-one interview with a trained health worker, where basic demographic information (age, education), past medical/ gynecologic history, family history, HPV knowledge assessment, and other behavioral factors (transportation method, access to medical care) were collected. The interview was conducted in the local Chinese dialect.

After the initial interview, a clinician collected a cervical exfoliated cellular (Pap) sample for HPV testing using a Qiagen cervical sampler brush (Qiagen) and a Whatman indicating FTA elute cartridge (GE Healthcare). Next, a second VIA examination was conducted, this time by the study clinician, and the location of any abnormal appearing lesion was recorded. Each patient then immediately underwent a standard colposcopic examination, conducted by the



Figure 1. A, design of the prescreening phase of the study. The study was designed to accrue an estimated 100 patients with cervical precancerous lesions and 100 normal patients. B, design of the HRME evaluation study. A total of 174 patients were recruited to the study from the prescreening phase.



99

100

101

140 same clinician, again with the location of any abnormal 141 appearing lesions recorded. Proflavine solution was then 142topically applied to the cervix (Fig. 2B). HRME imaging 143immediately followed, with gentle placement of the fiber 144 optic probe tip directly onto the site of interest (Fig. 2C). 145All colposcopically abnormal lesions were imaged with 146HRME, in addition to one colposcopically normal site per patient. Figure 2D shows the HRME image obtained with 147the probe placed at the site indicated in Fig. 2C. The imaged 148 field-of-view is a 720-µm wide en face view, corresponding 149 to the area of tissue beneath the diameter of the fiber 150optic probe tip. A biopsy was taken at each colposcopically 151 152abnormal site and immediately placed in fixative for stan-153dard histopathology processing. Colposcopic identification of lesions, HRME probe placement and imaging, and biopsy 154155collection were all conducted within a single examination, 156by the same clinician, in an attempt to co-register measure-157ment sites. Slides with hematoxylin and eosin (H&E)stained tissue sections were prepared at the Xiangyuan 158County Women's and Children's Hospital, China, and read 159by the study pathologist. The entire imaging portion of the 160 161 study was typically completed in 5 minutes.

162 HPV testing

163 Cervical specimens were also tested for 37 HPV genotypes
164 using the Roche HPV linear array test (Roche Diagnostics) at
165 Johns Hopkins University, as previously described (15).
166 Patients were considered positive for high-risk HPV (HR167 HPV) if their test was positive for any of the high-risk HPV

genotypes (16, 18, 31, 33, 35, 39, 45, 51, 52, 56, 58, 59, and/ 169 or 68). 170

171

178

Colposcopy

Biopsies were obtained from all lesions which were172visually apparent under colposcopy. Specimens were173processed for histopathology and graded by the study174pathologist as normal, cervical intraepithelial neoplasia175grade 1 (CIN1), grade 2 (CIN2), grade 3 (CIN3), or cancer176[squamous cell carcinoma (SCC)].177

High-resolution microendoscopy

Immediately before imaging with the HRME, topical 179proflavine solution (0.01% w/v in sterile PBS) was applied 180 to the cervix with a Q-tip, similar to application of acetic 181 acid in standard colposcopy. Proflavine is a fluorescent 182contrast agent which selectively stains cell nuclei. The dye 183 strongly absorbs blue light with an optical absorption peak 184 at a wavelength of 445 nm, producing green fluorescence 185emission with a peak wavelength of 515 nm. Once the dye 186was applied, patients immediately underwent HRME imag-187ing; no additional incubation period was necessary. Tech-188 nical details on the HRME design and assembly have been 189 described previously by Pierce and colleagues (14). Briefly, 190 the system operates as a compact, battery-powered fluores-191cence microscope, coupled to a flexible fiber optic imaging 192probe, 1 mm in diameter (Fig. 2A). Blue light provided by a 193light-emitting diode (LED) at a wavelength of 455 nm is 194delivered from the HRME unit, through the fiber optic 195



Figure 2. A, photograph of the HRME system. B–D, demonstration of the imaging procedure. B, proflavine is applied using a cotton-tipped swab. C, the fiber optic probe is placed in gentle contact with the cervix. D, a high-resolution image is displayed on a laptop computer in real-time. E, colposcopic view of an acetowhite cervical lesion at 5 o'clock (arrow). F, resulting HRME image. Histologic diagnosis of this site was normal, consistent with the HRME image which shows small, evenly spaced nuclei. G, colposcopic view of another acetowhite cervical lesion at 12 o'clock (arrow). H, resulting HRME image. Histologic diagnosis of this site was CIN3, consistent with the HRME image which shows large, crowded, pleomorphic nuclei. HRME image scale bars = 100 μm.

198 probe, to the tissue surface. Fluorescence from proflavine-199stained epithelium is transmitted back through the same 200probe to the HRME unit and imaged onto a CCD camera. 201 Images are displayed on a laptop computer screen in real-202time at 12 frames per second. The fiber optic probe used in 203the current study provides a 0.72-mm diameter field-of-204view with 4.4-µm resolution. After imaging each patient, the 205fiber optic probe was disinfected with Cidex OPA, according 206 to the manufacturer's instructions (Johnson & Johnson).

207**Data analysis**

234

235

236

237

HRME images of proflavine-stained tissue primarily 208209reveal cell nuclei as discrete bright dots on a dark back-210ground. To quantify parameters related to nuclear morphology, image analysis software was written (Matlab, 211212R2010b) to automatically identify nuclei, based on their 213characteristic size, shape, and brightness in HRME images. 214Raw grayscale images were subject to an adaptive histogram 215equalization algorithm to optimize contrast across the 216entire field-of-view, followed by a 2-dimensional median 217filter to reduce the appearance of the fiber optic probe's 218internal structure. A binary image was then generated by 219applying a single user-defined intensity threshold to a userselected region of interest in each image, leaving pixels with 220221original values above the threshold as 1, and pixels below 222the threshold as 0. Morphologic processing then removed 223 small objects (noise) and large objects (clumps) before 224labeling each group of connected "1" pixels as unique 225objects and documenting their properties (location, size, 226outline). Each object was considered to be an individual cell 227nucleus. The average nuclear-to-cytoplasm ("N/C") ratio 228for each image was calculated by dividing the total number 229of image pixels identified as nuclei, by the total number of 230pixels (minus nuclei and eliminated clumps) within the region of interest. Examples of this image processing pro-231232cedure are shown in Supplementary Fig. S1. 233

Before quantitative image analysis, each HRME image underwent a quality control (QC) review by one of the study investigators (M.C. Pierce), which removed images from the data pool if any of the following criteria were met: (i) The focused portion of the HRME image occupied less than half of the available field of view. (ii) there was excessive loose tissue or debris in the field of view, or (iii) there was cellular material/debris visibly adhered to the 241fiber tip. 242

Results

Visually apparent lesions were noted both by VIA and colposcopy in 62 patients, whereas VIA and colposcopic examinations were negative in 111 patients. No VIA or colposcopic impression was recorded for one patient and the HRME data obtained from this patient were not analyzed further. One hundred and ninety-five unique cervical sites were imaged in the remaining 173 patients. Seventynine of these sites were at colposcopically abnormal lesions in 62 patients. The remaining sites were at colposcopically normal locations in the remaining 111 patients. Figure 2 presents the colposcopic appearance and HRME images from abnormal appearing sites in 2 different patients. In Fig. 2E, the acetowhite region at 5 o'clock was considered abnormal by the colposcopist. Following placement of the fiber optic probe onto this site, HRME imaging revealed nuclei appearing as discrete dots, sparsely and evenly distributed throughout the field-of-view, characteristic of normal squamous epithelium (Fig. 2F). Following biopsy, the pathology diagnosis for this site was non-neoplastic. A second patient with a lesion considered abnormal under standard colposcopy at the 12 o'clock location (Fig. 2G) also underwent HRME imaging. In the HRME image (Fig. 2H), nuclei appear more crowded and unevenly spaced, with some loose debris and mucus within the field-of-view. The histopathologic diagnosis at this site was CIN3.

We calculated N/C ratio values for each of the 69 colposcopically abnormal sites which passed the quality control review (10 sites were eliminated by QC review). These sites were biopsied on the basis of colposcopic appearance, therefore enabling comparison of HRME-derived N/C ratio values against the histopathologic diagnosis. We also established the N/C ratio at one imaged site in each of 95 colposcopically normal patients (images from 16 of these 111 patients were eliminated by QC). These sites were not biopsied and



Figure 3. A, individual N/C ratio values measured at each of the 69 sites with a pathology diagnosis, as well as the 95 colposcopically normal sites imaged. The dashed line represents a post hoc threshold at the lowest value which correctly classified all 12 CIN2+ sites as "neoplastic." B. mean SE values for N/C ratios at each pathology grade.

243

244

245

246

247

248

249

250

251

252

253

254

255

256

257

258

259

260

261

262

263

264

265

266

267

268

269

270

271

272

273

274

275

276

277

280therefore only permit comparison of N/C ratio to colpo-281scopic appearance. Figure 3A shows the individual N/C ratio 282values measured at each of the 69 sites with a pathology 283diagnosis as well as the 95 colposcopically normal sites 284imaged. Figure 3B shows the mean SE values for N/C 285ratios at each pathology grade. The 12 lesions diagnosed as 286CIN2/3/SCC had higher mean and median N/C ratios than each of the lower grade categories. Combining all diagnoses 287288less severe than CIN2 into a single category (median N/C =2890.164) and comparing with the CIN2 or more severe diagnosis (median N/C = 0.251) indicated that the mean ranks of 290291 the N/C ratio values are significantly different for these 2 292groups (Kruskal–Wallis, $P = 1.3601 \quad 10^{-4}$).

293Figure 3A also shows a horizontal dashed line representing a post hoc N/C ratio threshold value of 0.185, which was 294295the lowest value that correctly classified all 12 CIN2+ sites 296as "neoplastic," Using this threshold, 38 of 57 sites consid-297 ered abnormal by colposcopy (and therefore biopsied) were 298correctly classified as non-neoplastic by HRME, based on 299their histopathologic diagnoses (open markers below the threshold in Fig. 3A). Nineteen of these 57 sites with a 300 pathology grade of normal or CIN1 were incorrectly clas-301 sified by HRME as abnormal (open markers above the N/C 302 303 = 0.185 threshold in Fig. 3A). When the same threshold was 304 applied to the colposcopically normal (and not biopsied) 305 sites (Fig. 3A), only 8 of the 95 sites exhibited an N/C figure 306 above 0.185.

307 Table 1 summarizes the fraction of sites classified as 308 neoplastic by HRME versus histologic diagnosis, also strat-309 ified by colposcopic impression. Only 8.4% of sites with a 310normal colposcopic impression were classified as neoplastic 311 by HRME. All histologically neoplastic sites (CIN2+) were identified correctly by HRME. Of the 54 colposcopically 312313 abnormal sites with benign histology (false positive by colposcopy), only 35% were classified as neoplastic by 314 315HRME.

316 Table 2 summarizes the fraction of patients classified as 317 neoplastic by HRME versus histologic diagnosis, this time 318 stratified by whether the patient tested positive for high-risk HPV. Nine patients had histologically confirmed disease 319320 (CIN2+); all 9 tested positive for HR-HPV and also had an 321 N/C ratio above the 0.185 threshold on HRME. Among 322 women who were colposcopically normal or had a pathol-323 ogy diagnosis of <CIN2, there was no difference in the

percentage that had a HRME N/C ratio above 0.185 for
those who were HR-HPV negative (19 of 133 = 14.3%) and
those who were HR-HPV positive (6 of 32 = 18.8%; P = 0.6,
Fisher exact test).326
327

329

359

360

361

362

363

364

365

366

Discussion

Through improvements in our knowledge of the patho-330 genesis of cervical cancer, the disease is now mostly pre-331 ventable but still disproportionately affects women living in 332 developing countries. Screening and treatment approaches 333 based on cervical cytology have been successful in reducing 334 the burden of cervical cancer where effective programs have 335 been established. However, attempts to establish such pro-336 grams in lower resource settings, which experience more 337 than 80% of the burden of cervical cancer incidence and 338 mortality, have been largely unsuccessful. Several new tools 339 have emerged that may help to address these disparities, 340 including HPV vaccination, lower cost HPV DNA testing, 341 visual inspection methods, and ablative treatment (16). The 342 use of VIA with cryotherapy has enabled see-and-treat 343 programs to be implemented in several countries, providing 344 women with the opportunity for cervical screening and 345 treatment to be completed in a single clinic visit. However, 346 there remains concern about the real possibility of signif-347 icant overtreatment based on VIA-positive results, with a 348 similar concern for management of HPV-positive women 349participating in HPV test-based screening. This study eval-350 uated a recently developed low-cost imaging device which 351provides real-time information on cervical cell morphology 352 in vivo. Such information may prove complementary to 353 existing and emerging tools for diagnosis of cervical cancer 354and precancerous lesions in low-resource settings. Our 355 primary goal in this study was to evaluate whether HRME 356 imaging could potentially be used to improve the specificity 357 of visual inspection using either colposcopy or VIA. 358

Visually apparent lesions with a pathology diagnosis of CIN2 or higher exhibited more crowded nuclei, often with greater variation in nuclear size and separation than at sites graded as CIN1 or normal/benign. A *post hoc* single threshold value of N/C area ratio discriminated between sites with non-neoplastic and neoplastic pathology with 100% sensitivity (12 of 12 with CIN2+) and 67% specificity (38 of 57 with colposcopically positive lesions, but <CIN2 on

Table 1. Fraction of sites classified positive by HRME image analysis versus colposcopic impression and histologic diagnosis

Colposcopic impression	Histologic diagnosis	No. of sites measured	No. of sites HRME positive	% Sites HRME positive
Normal	No biopsy	95	8	8.4
Abnormal	Normal/benign	54	19	35
	CIN1	3	0	0
	CIN2	7	7	100
	CIN3	3	3	100
	SCC	2	2	100

High-risk HPV status	Histologic diagnosis	No. of patients measured	No. of patients HRME positive	% Patients HRME positive
HR HPV	No biopsy	90	5	5.6
	Normal/benign	43	14	32.6
HR HPV ⁺	No biopsy	21	2	9.5
	Normal/benign	9	4	44.4
	CIN1	2	0	0
	CIN2	6	6	100
	CIN3	2	2	100
	SCC	1	1	100

Table 2. Fraction of sites classified positive by HRME image analysis versus high-risk HPV test status and histologic diagnosis

369 pathology). Significantly, the 57 pathologically non-neo-370 plastic sites were all deemed sufficiently abnormal in appearance under VIA/colposcopy to warrant biopsy col-371372 lection, a false-positive rate of 57/69 = 83%. More than two 373 thirds of those unnecessary biopsies were identified as non-374neoplastic by HRME imaging. Of the 19 colposcopically abnormal sites which were incorrectly classified by HRME as 375376 abnormal against a gold-standard of pathology, the major-377 ity of these false-positive sites (17 of 19) showed chronic 378 inflammation, either alone or with metaplasia (Fig. 3A). 379Given the generally high prevalence of inflammation in 380 patients in low-resource settings, such conditions may 381 impact the accuracy of HRME in these populations. How-382 ever, the results reported here from China suggest that 383 HRME may improve specificity over VIA alone while 384emphasizing the need for further evaluation in populations 385with even higher prevalence of inflammation.

When patients were initially stratified based on a positive 386 high-risk HPV DNA test, HRME image analysis correctly 387 identified 100% of patients with CIN2 or more severe 388 389disease (9 of 9 patients). Of the 30 patients with a positive 390high-risk HPV test but no histologic (11 patients < CIN2) or 391colposcopic (21 patients) evidence of disease, only 6 392patients (18.8%) were identified as neoplastic by HRME 393imaging. These data support the potential for HMRE imag-394 ing to be used as an adjunct diagnostic tool in settings where 395 HPV testing provides the initial screening result.

396 While reducing the amount of unnecessary biopsies can 397 reduce program costs, the HRME system used in this study 398 requires an upfront investment of around \$3,000, the majority of this cost being allocated to the imaging camera. 399 400 We have also evaluated lower cost consumer-grade cameras 401 that retail for around \$300 and have confirmed their suit-402ability for use in HRME system (17). HRME is not the first in 403vivo cellular-level imaging technique to be evaluated for 404 detecting cervical neoplasia. The use of confocal microsco-405py, in reflectance and fluorescence modes, has been 406 reported previously with promising results. The study by 407 Tan and colleagues (13) showed the ability of confocal 408 fluorescence microscopy to also visualize nuclear morphology in the cervical epithelium following topical application 410 of acriflavine dye (proflavine, the dye used in the study 411 described here, is the fluorescent component of acriflavine). 412 The authors developed a set of qualitative criteria which 413 readers could apply to each image to assist in reaching a 414 diagnostic decision. In a prospective study of 15 tissue sites, 415each with an independent pathology diagnosis, readers 416 achieved 97% sensitivity and 93% specificity in identifying 417 sites classified by pathology as CIN2 or higher. We note that 418 the descriptive criteria developed by Tan and colleagues 419were entirely based on features related to nuclear morphol-420ogy and could be directly applied to images generated by the 421 HRME system in real-time. HRME images also display an 422area of tissue 2.5 times larger at a frame rate 6 times faster 423than that of the confocal platform used by Tan and collea-424gues, which may also improve diagnostic performance and 425ease of use. An assessment of training methods and learning 426curve for users of HRME was not directly included within 427 this study, although it may be noted that all image data were 428acquired by clinical staff with no prior experience of HRME 429imaging. We have found that HRME images can be accu-430rately classified as neoplastic/non-neoplastic by clinicians 431 using qualitative criteria in other organ sites (18, 19). 432However, we showed here the ability to objectively classify 433images using quantitative analysis of features such as N/C 434 ratio, thereby reducing the degree of subjective interpreta-435tion required by the user. Future studies will more thor-436 oughly assess these important questions which will impact 437 uptake of this technology. 438

While the HPV vaccine has immense potential to positively impact cancer prevention programs, particularly in low-resource settings, early detection techniques will still have key roles to play. The cost of the vaccine is declining but may remain unacceptably high in some regions. Current HPV vaccines target the 2 types, HPV16 and 18, which are only responsible for 70% of cervical cancers. Even when a suitably priced vaccine becomes widely available and is given to adolescents, it will take decades for the impact of vaccination to become clear. Women who have already been exposed to HPV and will not benefit from HPV

439

440

441

442

443

444

445

446

447

448

449

452 vaccination, and those who are have not been vaccinated
453 will still require screening, ideally at between 30 and 45
454 years of age (20, 21).

The limitations of this study include a design which did not exactly reflect the intended use of the HRME in practice (i.e., as an adjunctive diagnostic tool for use in triaging VIA⁺ or HPV⁺ cases). However, this pilot study was carried out to permit evaluation of a relatively large VIA and HPV

460 population that will certainly be encountered in the field.

- 461 These data will require validation in a larger, well-powered,
- 462 prospective trial that permits objective evaluation and 463 establishment of HRME image features, including the opti-
- 464 mum N/C ratio threshold for classifying tissue as neoplastic.

465 **Disclosure of Potential Conflicts of Interest**

466R. Richards-Kortum serves as an unpaid scientific advisor to Remicalm467LLC, holds patents related to optical diagnostic technologies that have been468licensed to Remicalm LLC, and holds minority ownership in Remicalm LLC.469P. Castle has been compensated by Merck for serving on a Data and Safety470Monitoring Board for HPV vaccines and has received HPV tests and testing for471research at a reduced rate or no cost from Qiagen and Roche. No potential472^{Q3}conflicts of interest were disclosed by the other authors.

473 Authors' Contributions

474^{Q4} **Conception and design:** M.C. Pierce, P. Castle, R. Richards-Kortum

501 **References**

502

503

504

505

506

507

508

509

510

511

512 513

514

515

516

517

518

519

520

521

522 523

 $524 \\ 525$

526

527

528

529

530

531

532

533

- Ferlay J, Shin HR, Bray F, Forman D, Mathers C, Parkin DM. GLO-BOCAN 2008 v1.2, Cancer Incidence and Mortality Worldwide: IARC Cancer Base No. 10. Lyon, France: International Agency for Research on Cancer; 2010. [cited 2012 Jan 17]; Available from: http://globocan. iarc.fr.
 - 2. Boyle P, Levin B, editors. World Cancer Report 2008. Lyon, France: International Agency for Research on Cancer; 2008.
 - 3. Sankaranarayanan R, Gaffikin L, Jacob M, Sellors J, Robles S. A critical assessment of screening methods for cervical neoplasia. Int J Gynecol Obstet 2005;89:Suppl 2:S4–12.
 - Denny L, Kuhn L, Pollack A, Wright TC Jr. Direct visual inspection for cervical cancer screening: an analysis of factors influencing test performance. Cancer 2002;94:1699–707.
 - Sauvaget C, Fayette J-M, Muwonge R, Wesley R, Sankaranarayanan R. Accuracy of visual inspection with acetic acid for cervical cancer screening. Int J Gynaecol Obstet 2011;113:14–24.
 - Goldie SJ, Gaffikin L, Goldhaber-Fiebert JD, Gordillo-Tobar A, Levin C, Mahe C, et al. Cost-effectiveness of cervical-cancer screening in five developing countries. N Engl J Med 2005;353:2158–68.
 - Cronje HS, Parham GP, Cooreman BF, de Beer A, Divall P, Bam RH. A comparison of four screening methods for cervical neoplasia in a developing country. Am J Obstet Gynecol 2003;188:395–400.
 - Thekkek N, Richards-Kortum R. Optical imaging for cervical cancer detection: solutions for a continuing global problem. Nat Rev Cancer 2008;8:725–31.
 - Georgakoudi I, Sheets EE, Müller MG, Backman V, Crum CP, Badizadegan K, et al. Trimodal spectroscopy for the detection and characterization of cervical precancers *in vivo*. Am J Obstet Gynecol 2002;186:374–82.
- Chang VT-C, Cartwright PS, Bean SM, Palmer GM, Bentley RC, Ramanujam N. Quantitative physiology of the precancerous cervix *in vivo* through optical spectroscopy. Neoplasia 2009;11:325–32.

Development of methodology: M.C. Pierce, R. Richards-Kortum	476
Acquisition of data (provided animals, acquired and managed patients,	477
provided facilities, etc.): Y.Y. Guan, WH. Zhang, YL. Qiao	478
Analysis and interpretation of data (e.g., statistical analysis, biosta-	479
tistics, computational analysis): M.C. Pierce, M.K. Quinn, P. Castle, R.	480
Richards-Kortum	481
Writing, review, and/or revision of the manuscript: M.C. Pierce, Y.Y.	482
Guan, M.K. Quinn, P. Castle, R. Richards-Kortum	483
Administrative, technical, or material support (i.e., reporting or orga-	484
nizing data, constructing databases): Y.Y. Guan, X. Zhang, YL. Qiao	485
Study supervision: Y.Y. Guan, R. Richards-Kortum	486

Acknowledgments

The authors thank the contribution of Dr. Patti Gravitt of the Johns Hopkins School of Public Health, in processing and evaluating the HPV tests. They also thank the contributions of Shaoming Wang and Changyan Feng in patient pre-screening and registration and preliminary work by Lina Hu and Deepika Satish on data analysis. 487

488

489

490

491

492

493

494

495

496

497

498

535

536

537

538

539

540

541

542

543

544

545

546

547

548

549

550

551

552

553

554

555

556

557

558

559

560

561

562

563

564

565

566

Grant Support

This research was funded by the U.S. NIH grant R01 EB007594. The costs of publication of this article were defrayed in part by the payment of page charges. This article must therefore be hereby marked *advertisement* in accordance with 18 U.S.C. Section 1734 solely to indicate this fact.

Received May 25, 2012; revised July 6, 2012; accepted July 27, 2012; 499 published OnlineFirst xx xx, xxxx. 500

- Pierce MC, Javier DJ, Richards-Kortum R. Optical contrast agents and imaging systems for detection and diagnosis of cancer. Int J Cancer 2008;123:1979–90.
- Sung K-B, Richards-Kortum R, Follen M, Malpica A, Liang C, Descour MR. Fiber optic confocal reflectance microscopy: A new real-time technique to view nuclear morphology in cervical squamous epithelium *in vivo*. Opt Express 2003;11:3171–81.
- Tan J, Quinn MA, Pyman JM, Delaney PM, McLaren WJ. Detection of cervical intraepithelial neoplasia *in vivo* using confocal endomicroscopy. BJOG 2009;116:1663–70.
- **14.** Pierce MC, Yu D, Richards-Kortum R. High-resolution fiber-optic microendoscopy for *in situ* cellular imaging. JOVE 2011;47:e2306.
- Gravitt PE, Coutlee F, Iftner T, Sellors JW, Quint WGV, Wheeler CM. New technologies in cervical cancer screening. Vaccine 2008;26S: K42-K52.
- Schiffman M, Castle PE. The promise of global cervical-cancer prevention. N Engl J Med 2005;353:2101–4.
- Shin D, Pierce MC, Gillenwater AM, Williams MD, Richards-Kortum RR. A fiber-optic fluorescence microscope using a consumer-grade digital camera for *in vivo* cellular imaging. PLoS One 2010;5:e11218.
- Muldoon TJ, Thekkek N, Roblyer D, Maru D, Harpaz N, Potack J, et al. Evaluation of quantitative image analysis criteria for the high-resolution microendoscopic detection of neoplasia in Barrett's esophagus. J Biomed Opt 2010;15:026027.
- Muldoon TJ, Roblyer D, Williams MD, Stepanek VMT, Richards-Kortum R, Gillenwater AM. Noninvasive imaging of oral neoplasia with a high-resolution fiber-optic microendoscope. Head Neck 2012;34: 305–12.
- Agosti JM, Goldie SJ. Introducing HPV vaccine in developing countries -Key challenges and issues. N Engl J Med 2007;356:1908–10.
- 21. Roden R, Wu T-C. How will HPV vaccines affect cervical cancer? Nat Rev Cancer 2006;6:753–63.